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2 **An actin-based viscoplastic lock ensures progressive body**

3 **axis elongation**

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23 **Abstract**

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A key step in animal development is the process of body axis elongation, laying out the
27 final form of the entire animal. This critically depends on polarized cell shape changes¹,
28 which rely on the interplay between intrinsic forces generated by molecular motors²⁻⁵,
29 extrinsic forces due to adjacent cells pulling or pushing on the tissue⁶⁻⁹, and mechanical
30 resistance forces due to cell and tissue elasticity or friction¹⁰⁻¹². Understanding how
31 mechanical forces influence morphogenesis at the cellular and molecular level remains
32 a critical challenge². Recent work outlined that cell shape changes occur through small
33 incremental steps^{2,4,5,13}, suggesting the existence of specific mechanisms to stabilize cell
34 shapes and counteract cell elasticity. Here, we identify a spectrin-kinase-formin network
35 required to stabilize embryo shape when repeated muscle contractions promote *C.*
36 *elegans* embryo axis elongation. Its absence induces complete axis retraction due to
37 damage of epidermal actin stress fibers. Modeling predicts that a mechanical
38 viscoplastic deformation process can account for embryo shape stabilization. Molecular
39 analysis suggests that the physical basis for viscoplasticity originates from the progressive
40 shortening of epidermal microfilaments induced by muscle contractions and FHOD-1
41 formin activity. Our work thus identifies an essential molecular lock acting in a
42 developmental ratchet-like process.

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46 *C. elegans* provides a simple and integrated model to study the cellular impact of
47 mechanical forces. Its embryonic elongation only relies on cell shape changes and
48 includes two phases depending on tension and stiffness anisotropies in the epidermis¹²,
49 and beyond the 2-fold stage on muscle activity⁸ (Fig. 1a; Supplementary material).
50 Importantly, neither stage relies on pulsatile actomyosin flows¹², as observed in the early
51 *C. elegans* zygote, or during *Drosophila* gastrulation and germband extension^{2,4,5,13}.
52 Because muscles are tightly mechanically coupled to the epidermis through epidermal
53 hemidesmosomes¹⁴, their contractions also displace the epidermis. This can be
54 monitored by tracking the anterior-posterior displacement of muscle nuclei and the
55 circumferentially oriented epidermal actin filaments (Fig. 1b-b'''). Importantly, all
56 muscles do not contract simultaneously (X. Yang and M. Labouesse, unpublished).
57 Hence, when some areas of the epidermis are longitudinally compressed (red line in Fig.
58 1c'), others are stretched (green line in Fig. 1c') before eventually relaxing (Fig. 1c-c'';
59 Fig. S1). Importantly, we previously established that the tension generated by muscle
60 activity triggers a mechanotransduction pathway in the epidermis, which promotes axis
61 elongation⁸.

62 The relaxation observed after each muscle contraction raises a conundrum: how can
63 muscle activity power embryonic elongation from 100 μm to 200 μm within an hour if cell
64 elasticity brings cells back to their initial length after each contraction (Fig. 1d). One
65 simple hypothesis would be that some mechanism stabilizes the transient body
66 deformations induced by muscle activity (Fig. 1d'), as was proposed for *Drosophila*
67 gastrulation or germband extension in processes involving myosin II^{5,15,16}. To uncover this
68 mechanism, we focused on the kinase PAK-1, which lies at the crossroads of

69 hemidesmosome remodelling⁹ and actomyosin force regulation^{17,18}. We first performed
70 a feeding RNAi screen in a strong yet viable *pak-1* mutant, looking for enhancers of
71 embryonic lethality and/or body morphology defects (Fig. 2a). This screen identified the
72 gene *spc-1* encoding α -spectrin as a strong *pak-1* genetic enhancer leading to short
73 hatchlings (58 μm on average), which were significantly shorter than *pak-1(tm403)* (178
74 μm) or *spc-1(RNAi)* (91 μm) hatchlings (Fig. 2b, Table S1). The study of this genetic
75 interaction was further motivated by the identification of the central Src Homology 3
76 domain (SH3) of SPC-1 as an interactor of the N-terminal domain of PAK-1 in a yeast
77 two-hybrid screen (Fig. 2c, Table S2). Both screens thus point to an interaction between
78 SPC-1 and PAK-1.

79 To understand why *pak-1(tm403) spc-1(RNAi)* are shorter than *spc-1(RNAi)* embryos, we
80 examined their elongation rate using DIC microscopy. Wild-type and *pak-1(tm403)*
81 embryos initially elongated at the same rate, whereas *spc-1* defective embryos
82 elongated slower and stopped around the 2-fold stage (100 μm) as previously
83 described¹⁹ (Fig. 2d). By contrast, *spc-1(ra409) pak-1(tm403)* and *spc-1(RNAi) pak-*
84 *1(tm403)* embryos reached ≈ 65 μm at a slow rate, but then could not maintain their
85 shape, retracting back to ≈ 50 μm , which neither *spc-1(ra409)* nor *pak-1(tm403)* embryos
86 did (Fig. 2d, Movie 1). Two observations suggest that this phenotype is linked to muscle
87 activity. First, *spc-1* knock-down in *git-1* or *pix-1* mutants, two other players involved in
88 the same mechanotransduction pathway as PAK-1⁹, also induced retraction (Fig. S2).
89 Second, *spc-1(RNAi) pak-1(tm403)* embryos started to retract at the onset of active
90 muscle contraction in control embryos (pink box in Fig. 2d). To directly prove this
91 hypothesis, we abrogated muscle function in *spc-1(ra409) pak-1(tm403)* embryos by
92 knocking-down the kindlin homolog UNC-112²⁰. Strikingly, *spc-1(ra409) pak-1(tm403)*

93 embryos defective for *unc-112* no longer retracted (Fig. 2e; Movie 2). We conclude that
94 the mechanical input provided by muscles to the epidermis induces the retraction
95 phenotype observed in *spc-1 pak-1* double mutants.

96 The simplest interpretation of the retraction phenotype described above is that a
97 cellular structure maintaining embryo shape fails to emerge or collapses in *spc-1 pak-1*
98 double mutants once muscles become active. Two arguments lead us to consider that
99 this structure corresponds to the actin cytoskeleton. First, SPC-1/ α -spectrin and its
100 binding partner SMA-1/ β -spectrin form an actin-binding hetero-tetramer colocalizing
101 with actin²¹ and partially with PAK-1 in epidermal cells (Fig. S3). Second and foremost, it
102 has long been known that treating *C. elegans* embryos with the actin-depolymerizing
103 drug cytochalasin-D induces a retraction phenotype very similar to that presented
104 herein²². We thus characterized actin filaments by spinning-disk confocal imaging of a
105 LifeAct::GFP probe¹². Segmentation analysis of the fluorescence signal associated with
106 actin filaments in the dorso-ventral epidermis (Fig. 3a-a''') revealed more discontinuity in
107 *spc-1 pak-1* double deficient embryos (Fig. 3d-d''') compared to the control genotypes
108 (Fig. 3a-c'''). Moreover, Fourier transform analysis indicated that their degree of
109 anisotropy relative to the circumferential axis was abnormal (Fig. 3d''-d'''). Note that
110 both phenotypes were visible mainly at mid elongation, i.e. after muscles become
111 active (Fig. S4), suggesting that a well-structured actin network emerges too slowly in
112 double mutants rather than it collapses.

113 Collectively, the results described so far, together with the retraction phenotype of
114 cytochalasin-D treated embryos²², suggest that the actin filament defects account for
115 *spc-1 pak-1* embryo retraction, and further link muscle activity with these defects.

116 Significantly, as the wild-type embryo lengthens, its circumference decreases by roughly
117 20% due to embryo volume conservation (Fig. S5) and thus, the length of actin filaments
118 in dorso-ventral cells should decrease. Hence, we suggest that muscle activity normally
119 promotes actin filament shortening, probably through sliding or shortening of filaments
120 relative to each other after their bending (Fig. S1). We further suggest that SPC-1 and
121 PAK-1 stabilize cell shape by maintaining actin bundle integrity. We could not define the
122 shortening mechanism by spinning-disk microscopy, probably because each muscle
123 contraction results in changes beyond the time and space resolution of the microscope.
124 However, we suggest that it goes awry in the absence of SPC-1 and PAK-1, due to the
125 lack of a capping or bundling activity (Fig. 4b-b').

126 To rationalize the role of muscles in the process of actin bundle shortening and
127 stabilization, we described the *C. elegans* embryo as a Kelvin-Voigt material (a spring in
128 parallel to a dashpot) submitted to forces acting in the epidermis and muscles (F_{epid} and
129 $F_{muscles}$) (Fig. 4c equations-a and -b; Supplementary material). Note that F_{epid} is written as
130 the product of an active force, F_{seam} , and a passive component resulting from actin
131 bundle stiffness, α_{DV} (Supplementary material and ref. 11). Since muscle-defective
132 mutants cannot elongate beyond the 2-fold stage, then F_{epid} can only extend embryos
133 until that stage (due to the spring restoring force; Fig. S6a-a'). Simply adding the force
134 $F_{muscles}$ should not trigger any further extension, because it oscillates between a positive
135 and negative input (Fig. 1bc, Fig. S6b-b').

136 Recently, several studies have suggested that systems exposed to a stress can undergo
137 a permanent rearrangement, which can be described as a plastic deformation²³ or as
138 a change in the spring resting length^{24,25}. Accordingly, we incorporated an increase of

139 the spring resting length λ in the equations described above by writing that it changes
140 by a factor β (Fig. 4c equation-c; Fig. 4d). Thereby, we could accurately predict the
141 elongation pattern of wild-type embryos (Fig. 4f, Fig. S6d-d'; Supplementary material).
142 Conversely, in *spc-1 pak-1* defective embryos, the continuing damage to actin
143 filaments should reduce their stiffness (component α_{DV} in Fig. 4c equation-b), which we
144 expressed by writing that it depends on a tearing factor γ (Fig. 4c equation-d, Fig. 4e);
145 thereby, we could accurately predict their retraction pattern (Fig. 4f, Fig. S6e-e'). We
146 thus propose that SPC-1/ α -spectrin and PAK-1 regulate a process of mechanical
147 plasticity in the physical sense. From a cellular standpoint, having a changing resting
148 length means that body elasticity does not bring the embryo back to its initial shape
149 upon muscle relaxation, enabling progressive lengthening.

150 To further define the molecular basis of viscoplasticity, we performed a small-scale RNAi
151 screen to search for gene knockdowns inducing retraction of *spc-1(ra409)* embryos (Fig.
152 4g; Table S3). This screen identified the atypical formin FHOD-1 (Fig. 4h-i; Fig. S7, movie
153 3), which has previously been linked to actin dynamics in the epidermis²⁶. We confirmed
154 that *fhod-1(tm2363); spc-1(RNAi)* embryos also showed a penetrant retraction
155 phenotype (Fig. 4h; Fig. S7). The identification of this specific formin was intriguing
156 because vertebrate FHOD1 promotes actin capping and bundling rather than
157 nucleation and elongation²⁷. It thus raised the tantalizing possibility that FHOD-1 activity
158 stabilizes the actin cytoskeleton while it gets remodeled under the influence of muscle
159 activity during embryo circumference reduction. Furthermore, the genetic interaction
160 suggests that FHOD-1 acts with SPC-1 and PAK-1. To examine this possibility, we tested
161 whether FHOD-1 derivatives removing at least the C-terminal DAD domain, predicted to
162 auto-inhibit formins²⁸, can rescue the retraction phenotype of *spc-1 pak-1* deficient

163 embryos. Strikingly, after epidermis-specific expression of a form lacking the FH2 and
164 DAD domains, transgenic *spc-1(RNAi) pak-1(tm403)* embryos did not retract and were
165 significantly longer than non-transgenic siblings; rescue was better than with the full-
166 length protein. By contrast, the DAD deleted form did not rescue and deletion of the
167 FH1-FH2-DAD domains marginally rescued retraction, arguing that the FH2 F-actin
168 nucleation domain is not necessary for rescue but that the FH1 is. Truncation of the C-
169 terminal DAD domain or of the FH1-FH2-DAD domains marginally rescued retraction (Fig.
170 4j). Importantly, an FH2-DAD truncation in the mammalian FHOD1 still enables it to
171 bundle actin²⁷, further strengthening the notion that FHOD-1 bundling activity is indeed
172 required and providing a potential molecular basis for viscoplasticity. It also indicates
173 that muscle-induced actin remodeling would primarily result from sliding and re-
174 bundling (see Fig. 4a-a'). Furthermore, we conclude that the retraction of *spc-1 pak-1*
175 deficient embryos mainly results from a lack of FHOD-1 activation.

176 Several factors could contribute to improperly regulate the activity of PAK-1 and FHOD-
177 1. First, SPC-1 could help recruit FHOD-1 and PAK-1, since FHOD-1::GFP and PAK-1::GFP
178 made small aggregates in SPC-1 defective embryos (Fig. S8). Second, we found that
179 the cycles of actin filament displacement induced by muscle contractions were almost
180 twice as short in *spc-1(RNAi) pak-1(tm403)* embryos compared to *pak-1(tm403)* and
181 wild-type controls (3 sec against 5.7 sec; Fig. S9, Movie 4). These shorter muscle
182 contractions might still induce actin filament shortening but not give enough time for
183 their stabilization. Third, PAK-1 might directly activate FHOD-1 downstream of the
184 mechanotransduction pathway induced by muscles, since *git-1* and *pix-1* mutations
185 combined with *spc-1* RNAi-knockdown also induced a retraction (see Fig. S2a-g).

186 Altogether, our data identify three proteins involved in stabilizing cell shapes in a system
187 involving two mechanically interacting tissues submitted to repeated stress. We propose
188 that the progressive shortening of actin filaments under the control of these factors
189 mediates a cellular viscoplastic process promoting axis elongation. A similar
190 spectrin/p21-activated kinase/FHOD1 system might operate in vertebrate tissues
191 comprising an epithelial layer surrounded by a contractile layer, such as our internal
192 organs. Interestingly, high FHOD1 expression correlates with poor prognosis of breast
193 cancer patients²⁹. Thus, a similar viscoplastic process might also influence the metastatic
194 properties of tumor cells positioned next to contractile cells.

195

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209

210 **AUTHORS' CONTRIBUTIONS**

211 ML conceived the project, GP initiated many aspects of the study while AL performed
212 most experiments, with contributions from TF for modelling, TF and JP for image analysis,
213 FL for the generation of FHOD-1 variants, CG shared data from a related screen, SK for
214 the *spc-1(ra409)* mini-RNAi screen, DR for technical assistance. ML wrote the manuscript
215 and all authors commented and proofread it (except SK, who was an intern), AL
216 assembled figures, TF wrote the Supplementary mathematical modelling material, and
217 FL the Materials section.

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220

221 **FIGURE LEGENDS**

222 **Figure 1: Muscle contractions deform the epidermis to their mechanical coupling**

223 **(a)** *C. elegans* embryonic elongation from comma to 2-fold stages depends on a
224 ROCK-promoted actomyosin force in seam cells (cyan) and actin-promoted stiffness in
225 dorso-ventral cells (orange); elongation beyond the 2-fold stage requires repeated
226 muscle contractions (red flash), which induce a PAK-1-dependent mechano-
227 transduction pathway. Open cross-sections (bottom) show muscle positions. **(b-b''')**
228 Epidermis actin filament (green) and muscle nucleus (red) tracking in a wild-type 2-fold
229 embryo. **(b')** Kymographs from the yellow rectangle area **(b)** showing the concurrent

230 displacement of epidermal actin and muscle nuclei. **(b'')** Resulting displacement
231 curves; **(b''')** quantification of the area between them; its low value underlines the tight
232 mechanical coupling between both tissues. Scale bar, 10 μm . **(c-c''')** A muscle
233 contraction/relaxation cycle illustrating its local impact on epidermal actin filaments in a
234 wild-type 2-fold embryo (timing in left corner). Yellow (relaxation), red (compression)
235 green (stretching) distances between landmarks denoted 1-4: **(c)** [1-2], 7.8 μm ; [2-3],
236 19.8 μm ; [3-4], 24.6 μm . **(c')** [1-2], 9.4 μm ; [2-3], 13.6 μm ; [3-4], 26.2 μm . **(c'')** [1-2], 8.0 μm ;
237 [2-3], 19.2 μm ; [3-4], 25.0 μm . In **(b-c)** the *Pepidermis* promoter is *Pdpy-7*. **(d)** Hysteresis
238 graph of an idealized elastic material returning to its initial shape after deformation
239 (top), or showing permanent deformation³⁰ (bottom).

240
241 **Figure 2: Loss of PAK-1 and SPC-1 triggers a muscle-dependent retraction of embryos**
242 **(a)** RNAi screen in a *pak-1* mutant identified *spc-1* as an enhancer (Table S1). **(b)** DIC
243 micrographs of newly hatched wild-type, *pak-1(tm403)* (scale bar: 10 μm), *spc-1(RNAi)*
244 and *spc-1(RNAi) pak-1(tm403)* (scale bar: 25 μm). Quantification of L1 hatchling body
245 length: wild-type (n=38); *pak-1(tm403)* (n=32); *spc-1(RNAi)*(n=26); *spc-1(RNAi) pak-*
246 *1(tm40)* (n=36). **(c)** A yeast two-hybrid screen using the PAK-1 N-term domain as a bait
247 identified the SPC-1 SH3 domain as a prey (orange background) (Table S2). **(d)**
248 Elongation profiles and corresponding terminal phenotypes of wild-type (n=5), *pak-*
249 *1(tm403)* (n= 5), *spc-1(RNAi)* (n=8), *spc(RNAi) pak-1(tm403)* (n=8). **(e)** Elongation profiles
250 in a muscle defective background. *unc-112(RNAi)* (n=5); *spc-1(RNAi)* (n=8); *unc-*
251 *112(RNAi)*; *pak-1(tm403) spc-1(ra409)* (n=5); *spc-1(RNAi) pak-1(tm403)* (n=8). Right
252 bracket **(d, e)**, extent of retraction for *spc-1(RNAi) pak-1(tm403)* embryos. Scale bars, 10
253 μm . Error bars, SEM.

254

255 **Figure 3: Actin filament defects in SPC-1 and PAK-1 defective embryos**

256 **(a-d)** Epidermal actin filaments visualized with the *Pdpy-7::LifeAct::GFP* reporter
 257 construct in wild-type **(a-a''')**, *pak-1(tm403)* **(b-b''')**, *spc-1(RNAi)* **(c-c''')** and *spc-1(RNAi)*
 258 *pak-1(tm403)* **(d-d''')** embryos at mid-elongation (2-fold equivalent) stage. Yellow
 259 rectangle, region of interest (ROI). Scale bar, 10 μ m. **(a'-d')** ROI after binarisation (green)
 260 and major axis detection (red), based on **(a''')** three steps of image treatment for
 261 continuity and orientation analysis. **(a''-d'')** Actin continuity: distribution of actin
 262 segments based on their length. Wild-type (n=16); *pak-1(tm403)* (n=21); *spc-*
 263 *1(RNAi)* (n=21); *spc-1(RNAi) pak-1(tm403)* (n=17) **(b'''-d''')** Actin filament orientation:
 264 the curves represent the number of actin filaments oriented perpendicular to the
 265 elongation axis (90° angle in wild-type) based on the Fast Fourier Transformation (FFT in
 266 a'''). Wild type (n=18); *pak-1(tm403)* (n=20); *spc-1(RNAi)* (n=18); *spc-1(RNAi) pak-*
 267 *1(tm403)* (n=18). Scale bars, 10 μ m (c, d, e, f), or 1 μ m (c', d', e', f'). P values, *<0,05;
 268 **<0,001; ***<0,0001; ns not significant.

269

270 **Figure 4: An actin-remodeling network providing mechanical plasticity ensures embryo**
 271 **elongation**

272 **(a-b')** Cellular model of embryo elongation. **(a-a')** In control embryos, muscle
 273 contractions (red arrows) provoke actin filament shortening in the dorso-ventral
 274 epidermis, probably through sliding or shortening, followed by SPC-1/PAK-1-dependent
 275 actin stabilization. Whether spectrin is found along (scenario 1) or between (scenario 2)
 276 actin filaments is unknown **(a)**. **(b-b')** In *spc-1 pak-1* deficient embryos, actin remodeling
 277 goes uncontrolled. **(c-f)** Viscoplastic mechanical model of embryo elongation. The
 278 embryo is represented as a Kelvin-Voigt solid (spring stiffness k , resting length λ , viscosity

279 η) submitted to the forces F_{epid} and F_{muscle} . System equations for the model. **(d)** Wild-type
280 case: an increasing resting length during stretching phases imparts mechanical
281 plasticity. **(e)** *spc-1 pak-1* mutants: F_{epid} progressively decreases. **(f)** Comparison of
282 experimental and predicted elongation curves taking the constitutive equations shown
283 in **(c)**. **(g)** A retraction screen in a *spc-1* mutant identifies *fhod-1*. **(h)** Snapshot at three
284 time-points of *spc-1* deficient embryos in control, *pak-1* or *fhod-1* backgrounds; **(i)**
285 terminal body length at hatching: *spc-1(ra409)* after feeding on L4440 control (n=21), or
286 *fhod-1(RNAi)* (n=25) bacteria. **(j)** *Pdpy-7* driven epidermis expression of truncated FHOD-
287 1 variants and terminal body length at hatching: *spc-1(RNAi)* (n=26); *spc-1(RNAi)pak-*
288 *1(tm403)* no transgene (n=36), FHOD-1(full length) (n=16), FHOD-1(Δ DAD) (n=17), FHOD-
289 1(Δ FH2-DAD) (n=38) and non-transgenic siblings (n=78), FHOD-1(Δ FH1-FH2-DAD) (n=18).
290 Scale bar, 15 μ m. Error bars, SEM. P values: * $<0,05$; ** $<0,001$; *** $<0,0001$; ns, not significant.

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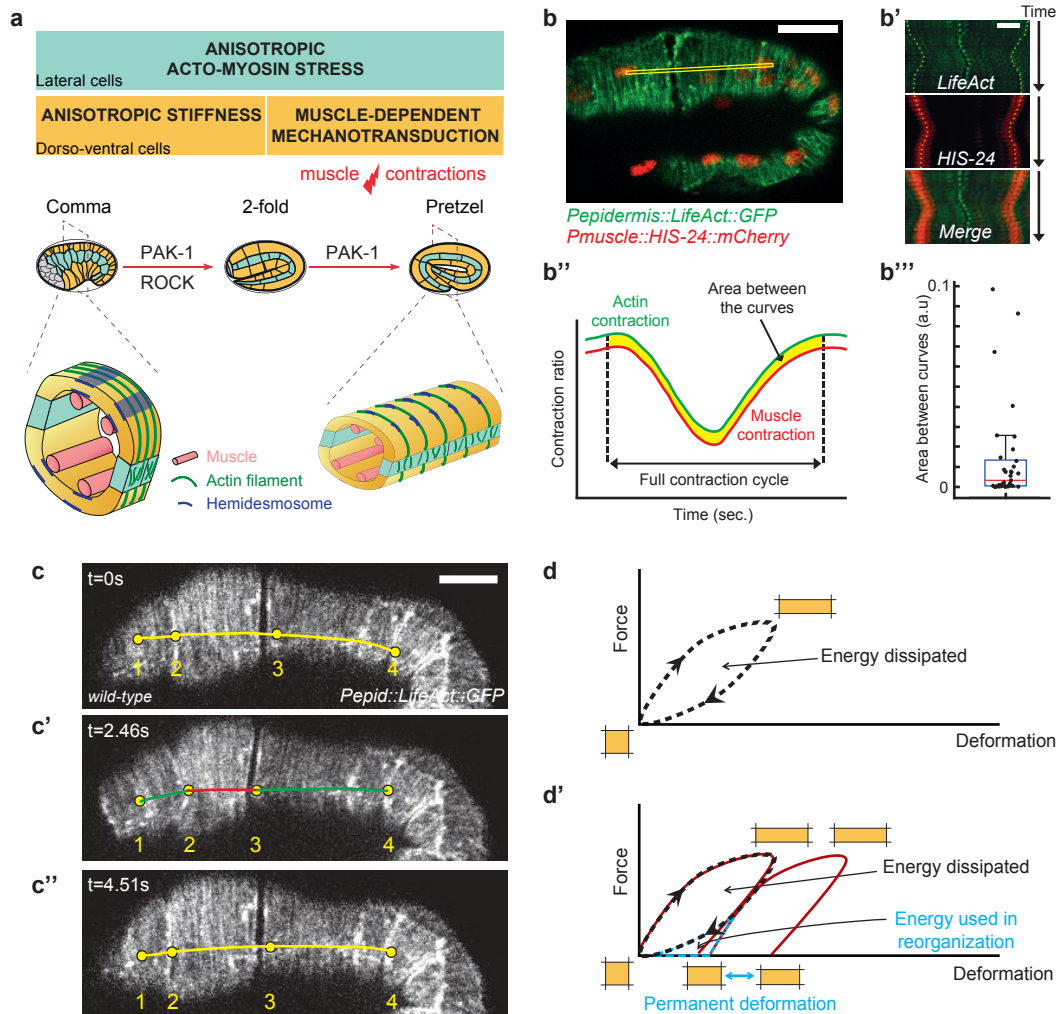


Figure 1: Muscle contractions deform the epidermis to their mechanical coupling

(a) *C. elegans* embryonic elongation from comma to 2-fold stages depends on a ROCK-promoted actomyosin force in seam cells (cyan) and actin-promoted stiffness in dorso-ventral cells (orange); elongation beyond the 2-fold stage requires repeated muscle contractions (red flash), which induce a PAK-1-dependent mechano-transduction pathway. Open cross-sections (bottom) show muscle positions.

(b-b''') Epidermis actin filament (green) and muscle nucleus (red) tracking in a wild-type 2-fold embryo.

(b') Kymographs from the yellow rectangle area (b) showing the concurrent displacement of epidermal actin and muscle nuclei.

(b'') Resulting displacement curves;

(b''') quantification of the area between them; its low value underlines the tight mechanical coupling between both tissues. Scale bar, 10 μm .

(c-c'') A muscle contraction/relaxation cycle illustrating its local impact on epidermal actin filaments in a wild-type 2-fold embryo (timing in left corner). Yellow (relaxation), red (compression) green (stretching) distances between landmarks denoted 1-4:

(c) [1-2], 7.8 μm ; [2-3], 19.8 μm ; [3-4], 24.6 μm .

(c') [1-2], 9.4 μm ; [2-3], 13.6 μm ; [3-4], 26.2 μm .

(c'') [1-2], 8.0 μm ; [2-3], 19.2 μm ; [3-4], 25.0 μm .

In (b-c) the *Pepidermis* promoter is *Pdpy-7*.

(d) Hysteresis graph of an idealized elastic material returning to its initial shape after deformation (top), or showing permanent deformation (bottom).

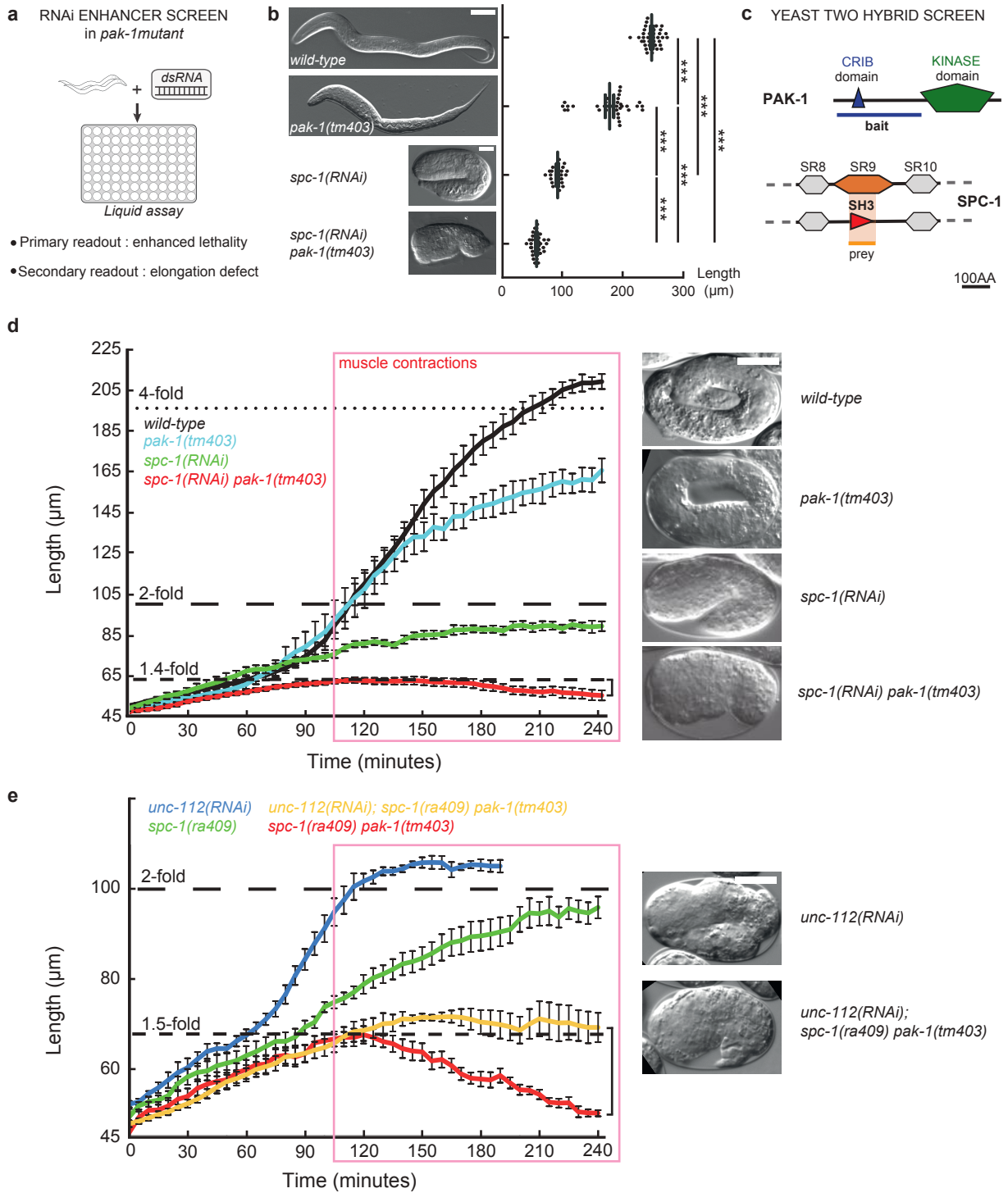


Figure 2: Loss of PAK-1 and SPC-1 triggers a muscle-dependent retraction of embryos

(a) RNAi screen in a *pak-1* mutant identified *spc-1* as an enhancer (Table S1).

(b) DIC micrographs of newly hatched wild-type, *pak-1(tm403)* (scale bar: 10 μm), *spc-1(RNAi)* and *spc-1(RNAi) pak-1(tm403)* (scale bar: 25 μm). Quantification of L1 hatching body length: wild-type (n=38); *pak-1(tm403)* (n=32); *spc-1(RNAi)*(n=26); *spc-1(RNAi) pak-1(tm403)* (n=36).

(c) A yeast two-hybrid screen using the PAK-1 N-term domain as a bait identified the SPC-1 SH3 domain as a prey (orange background) (Table S2).

(d) Elongation profiles and corresponding terminal phenotypes of wild-type (n=5), *pak-1(tm403)* (n=5), *spc-1(RNAi)* (n=8), *spc-1(RNAi) pak-1(tm403)* (n=8).

(e) Elongation profiles in a muscle defective background.

unc-112(RNAi) (n=5); *spc-1(RNAi)* (n=8); *unc-112(RNAi); pak-1(tm403) spc-1(ra409)* (n=5); *spc-1(RNAi) pak-1(tm403)* (n=8).

Right bracket (d, e), extent of retraction for *spc-1(RNAi) pak-1(tm403)* embryos. Scale bars, 10 μm . Error bars, SEM.

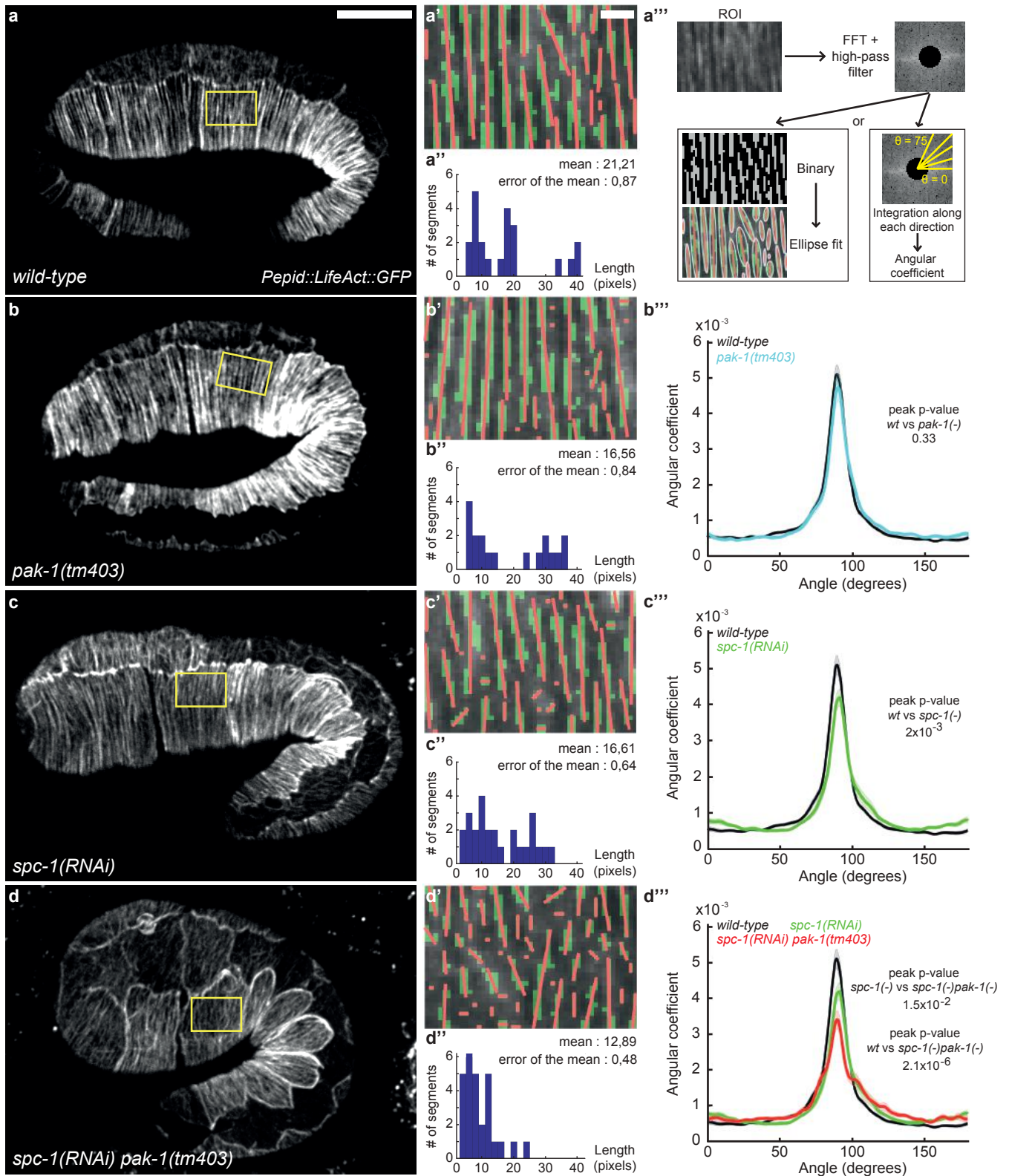


Figure 3: Actin filament defects in SPC-1 and PAK-1 defective embryos

(a-d) Epidermal actin filaments visualized with the *Pdp1::LifeAct::GFP* reporter construct in wild-type (a-a'''), *pak-1(tm403)* (b-b'''), *spc-1(RNAi)* (c-c''') and *spc-1(RNAi) pak-1(tm403)* (d-d''') embryos at mid-elongation (2-fold equivalent) stage. Yellow rectangle, region of interest (ROI). Scale bar, 10 μ m.

(a'-d') ROI after binarisation (green) and major axis detection (red), based on (a''') three steps of image treatment for continuity and orientation analysis.

(a''-d'') Actin continuity: distribution of actin

segments based on their length. Wild-type (n=16); *pak-1(tm403)* (n=21); *spc-1(RNAi)* (n=21); *spc-1(RNAi) pak-1(tm403)* (n=17)

(b''-d''') Actin filament orientation: the curves represent the number of actin filaments oriented perpendicular to the elongation axis (90° angle in wild-type) based on the Fast Fourier Transformation (FFT in a''').

Wild type (n=18); *pak-1(tm403)* (n=20); *spc-1(RNAi)* (n=18); *spc-1(RNAi) pak-1(tm403)* (n=18).

Scale bars, 10 μ m (c, d, e, f), or 1 μ m (c', d', e', f). P values, * $<0,05$; ** $<0,001$; *** $<0,0001$; ns not significant.

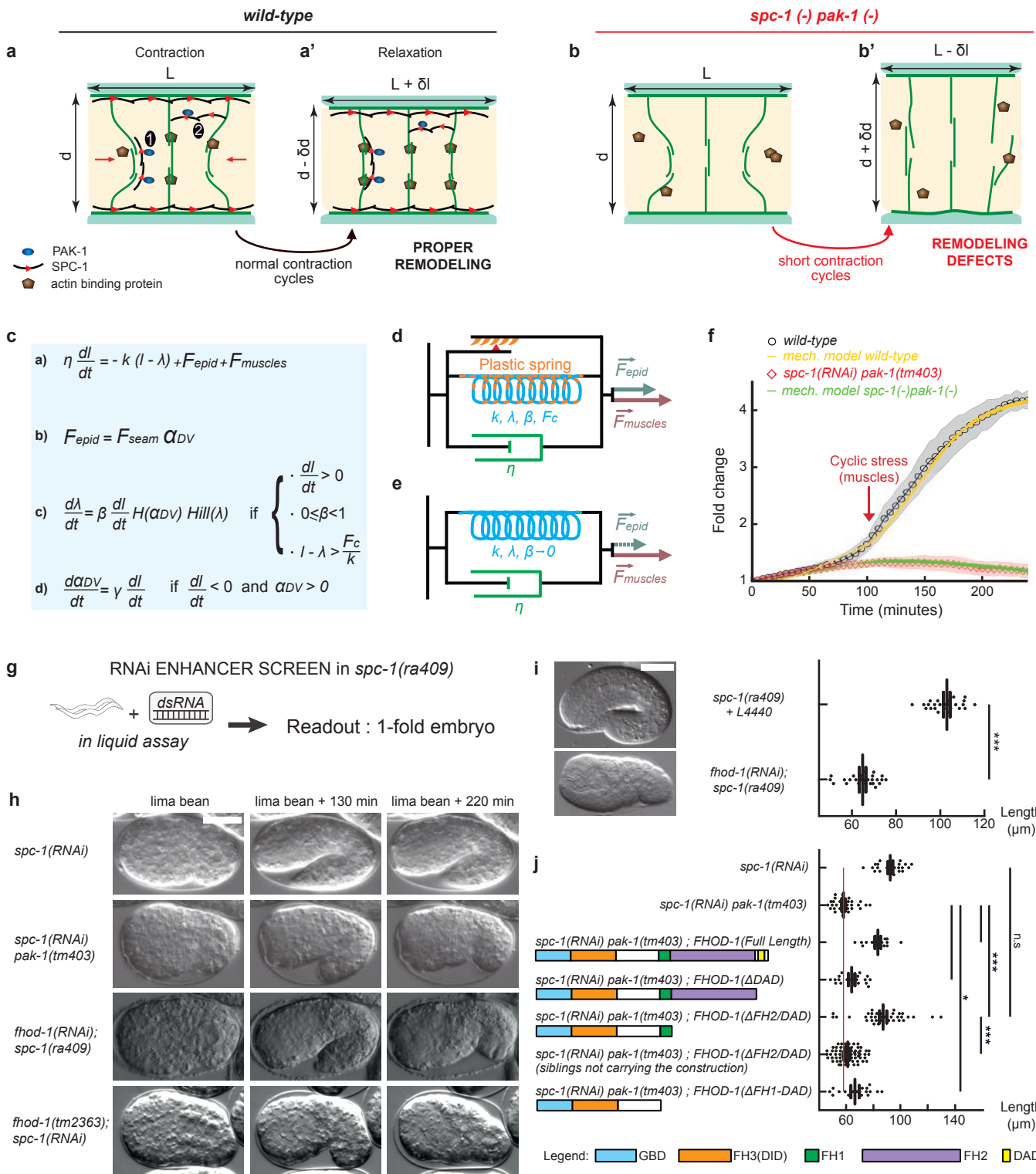


Figure 4: An actin-remodeling network providing mechanical plasticity ensures embryo elongation

(a-b') Cellular model of embryo elongation.

(a-a') In control embryos, muscle contractions (red arrows) provoke actin filament shortening in the dorso-ventral epidermis, probably through sliding or shortening, followed by SPC-1/PAK-1-dependent actin stabilization. Whether spectrin is found along (scenario 1) or between (scenario 2) actin filaments is unknown (a). (b-b') In *spc-1 pak-1* deficient embryos, actin remodeling goes uncontrolled.

(c-f) Viscoelastic mechanical model of embryo elongation. The embryo is represented as a Kelvin-Voigt solid (spring stiffness k , resting length λ , viscosity η) submitted to the forces F_{epid} and F_{muscle} . System equations for the model.

(d) Wild-type case: an increasing resting length during stretching phases imparts mechanical plasticity.

(e) *spc-1 pak-1* mutants: F_{epid} progressively decreases.

(f) Comparison of experimental and predicted elongation curves taking the constitutive equations shown in (c).

(g) A retraction screen in a *spc-1* mutant identifies *fhod-1*.

(h) Snapshot at three time-points of *spc-1* deficient embryos in control, *pak-1* or *fhod-1* backgrounds;

(i) terminal body length at hatching: *spc-1(ra409)* after feeding on L4440 control ($n=21$), or *fhod-1(RNAi)* ($n=25$) bacteria.

(j) Pdp-7 driven epidermis expression of truncated FHOD-1 variants and terminal body length at hatching: *spc-1(RNAi)* ($n=26$); *spc-1(RNAi) pak-1(tm403)* no transgene ($n=36$), FHOD-1(full length) ($n=16$), FHOD-1(Δ DAD) ($n=17$), FHOD-1(Δ FH2-DAD) ($n=38$) and non-transgenic siblings ($n=78$), FHOD-1(Δ FH1-FH2-DAD) ($n=18$). Scale bar, 15 μ m. Error bars, SEM. P values: * $<0,05$; ** $<0,001$; *** $<0,0001$; ns, not significant.

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SUPPLEMENTARY MATERIAL

An actin-based viscoplastic lock ensures progressive body axis elongation

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Content

- 1- Supplementary Mechanical Modeling
- 2- Methods
- 3- Supplementary Figures S1-S8 with legends
- 4- Captions for movies S1-S8
- 5- Supplementary References

1-Supplementary Mechanical Modeling

1.1 Background information. During embryogenesis the *C. elegans* embryo undergoes a process of elongation whereby it becomes four times as long as the long axis of the eggshell (50 μm). Cell proliferation and cell intercalation are absent, therefore the process of axis elongation relies only on the ability of the embryo to extend in the anterior-posterior direction. The outer epithelium (epidermis) plays an essential role in this process.

Changing the status of any physical entity requires the involvement of a force (mechanical or chemical), and the *C. elegans* embryo is no exception to this rule of physics. During the first phase of elongation and until muscles become active, the

24 machinery driving elongation involves an active force in the lateral epidermis (also
25 called seam cells), and a passive force exerted by the dorsal and ventral epidermal
26 cells (called DV cells) adjacent to the seam cells (Fig. 1a). Seam cells have a high
27 concentration of non-muscle myosin II, which has a non-polarized distribution and
28 does not display pulsatile flows^{1,2}, as observed for instance during *Drosophila*
29 germband elongation³. Nevertheless, the stress generated by the seam cells is
30 anisotropic and globally oriented along the DV axis (see cyan box in Fig. 1a)². The
31 stress anisotropy results mainly from the presence of circumferential F-actin filament
32 bundles in DV epidermal cells, which create a global stiffness anisotropy (see yellow
33 box in Fig. 1a). The DV epidermal cells do not contribute to generate active stress
34 (Fig. 1a), as their myosin II is kept mostly silent through the activity of the RhoGAP
35 RGA-2^{1,2,4}. The interplay between stress anisotropy in seam cells, stiffness anisotropy
36 from the DV epidermis, and hydrostatic pressure resulting from the reduction of
37 embryo diameter, induces a force oriented along the AP direction that is sufficient to
38 extend the embryo until it reaches the 2-fold stage².

39 Note that here as well as in the main text we refer to each elongation phase based
40 on the ratio between the actual embryo length and that of the eggshell long axis,
41 i.e. 1.7-fold or 2-fold means that the embryo has reached roughly 85 μm or 100 μm ,
42 respectively. Importantly, in mutant embryos which extend slower, we refer to
43 embryo stages based on the length that a wild-type embryo would reach after the
44 same time duration with t_0 corresponding to the beginning of elongation (see
45 extension curves in Fig. 2d and Movie 1).

46 For simplicity, let us call the net force in the AP direction produced by the epidermis
47 the epidermal cell force (F_{epid}). This force is not enough to explain the elongation up
48 to the 4-fold stage, since genetic analysis has established that embryos with non-

49 functional muscles do not elongate beyond the 2-fold stage (Fig. 2b)⁵. Therefore,
50 muscles provide a second active force driving elongation beyond the 2-fold stage,
51 which we will call $F_{muscles}$.

52 During embryogenesis, muscles organize and assemble in four rows located under
53 the epidermis (Fig. 1a). Muscles are attached to the extracellular matrix that
54 separates them from the epidermis, and that in turn serves to anchor the epidermis
55 through hemidesmosome-like junctions¹. Muscle organization and maturation is a
56 progressive process, such that muscle activity starts with small contractions at the 1.7-
57 fold, which progressively become more robust. The mechanical activity of muscles
58 can be summarized as an alternation of contractions followed by relaxation. Since
59 muscles are tightly connected to the epidermis, their contractions locally and
60 repeatedly induce an anterior-posterior compression and extension of the epidermis,
61 which can be visualized through the displacement of the actin cables (Fig. 1b-c, Fig.
62 S7). The stress exerted by muscle contractions on the epidermis induces a
63 mechanotransduction pathway (2nd yellow box in Fig. 1a), which is essential to
64 promote hemidesmosome maturation and embryo elongation⁸.

65

66 **1.2 Viscoplastic model.** The *C. elegans* epidermis can be modelled as a visco-elastic
67 body, more specifically as a Kelvin-Voigt system with a spring and dashpot in
68 parallel, subject to two main active forces: the epidermal force F_{epid} , which is a
69 continuous positive force, and the muscle force $F_{muscles}$, which is a pulsatile force since
70 muscles alternatively contract and relax. The first force is present since the beginning
71 of elongation, whereas the second force starts only after the 1.7-fold stage. The
72 elastic reaction of the epidermis to active forces can be captured by Hooke's law;
73 the damped nature of the reaction can be expressed by a viscous term. Overall the

74 length of the embryo over time $l(t)$ can be captured by the equation:

$$75 \quad \eta \frac{dl}{dt} = -k(l - \lambda) + F_{epid} + F_{muscles} \quad (1)$$

76 where k is the body stiffness, λ is the worm resting length and η is the coefficient of
77 viscosity. Inertia has been neglected given the low Reynolds number of the system.

78 Eq. (1) corresponds to the so-called Kelvin-Voigt viscoelastic model^{6,7} that captures
79 the behavior of viscoelastic solids under stress or deformation. For constant forces,
80 the solution of Eq. (1) is given by:

$$81 \quad l(t) = \frac{F_{epid} + F_{muscles}}{k} (1 - e^{-t/\tau}) + \lambda \quad (2)$$

82 meaning that the length of the system relaxes to the plateau value $\frac{F_{epid} + F_{muscles}}{k}$ in a
83 relaxation time of $\tau = \eta/k$.

84 F_{epid} can promote elongation until the 2-fold stage (Fig. S5a'). Beyond, the pulsatile
85 force originating from muscles, $F_{muscles}$, alternates periods of positive, negative or null
86 contribution, so that its temporal average $\langle F_{muscles} \rangle_{\theta}$ is null:

$$87 \quad \langle F_{muscles} \rangle_{\theta} = \frac{1}{\theta} \int_0^{\theta} F_{muscles}(t) dt = 0, \text{ for } \theta = nT \quad (3)$$

88 where θ is the integration period, T is the period and n is a positive integer. As a
89 consequence, on average, $F_{muscles}$ will not contribute to the steady state length (l_{ss})
90 of the embryo (see Fig. S5a-c) that is set by $l_{ss} = \frac{F_{epid}}{k} + \lambda$.

91 A way to introduce a positive contribution to embryo lengthening is to allow some
92 plasticity, in the physical sense, or ability of the system to get reorganized. For
93 example, let consider a stretching pulse due to muscle activity during which the
94 embryo increases its length $l(t)$ with an increment dl , such that the embryo will be

95 temporarily $l(t)+dl$ long at the end of the pulse. During the subsequent relaxation
 96 phase, due to elasticity in the system, the embryo should return to the initial length $l(t)$
 97 it had before muscles had locally extended it. However, if it undergoes a permanent
 98 plastic deformation, then the body will permanently keep a portion of the stretched
 99 length. This situation corresponds to a permanent rearrangement, and it has been
 100 observed and modeled in biological systems undergoing stresses^{8,9}. Similarly to⁹, a
 101 simple mathematical solution to introduce plasticity consists in having an adjustable
 102 resting length λ that increases linearly with the length $l(t)$ according to:

$$103 \quad \frac{d\lambda}{dt} = \beta \frac{dl}{dt} H(\alpha_{DV}) \quad \text{if } \frac{dl}{dt} > 0 \text{ and } l - \lambda > \frac{F_c}{k} \quad (4)$$

104 where $0 \leq \beta < 1$ is a proportionality factor called 'plasticity factor'; the case of $\beta = 0$
 105 corresponds to an absence of plasticity. The condition $\frac{dl}{dt} > 0$ ensures that there is
 106 rearrangement only during the extension phases and the condition; $l - \lambda > \frac{F_c}{k}$
 107 means that the rearrangement takes place only if the applied force exceeds a
 108 critical force F_c . The term $H(\alpha_{DV})$ is the Heaviside step function, which expresses that
 109 dorso-ventral rearrangement is possible only in presence of resistance. For the
 110 description of α_{DV} term, see next paragraph. Note that Eq. (4) is equivalent to
 111 $\lambda = \lambda(0) (1 - \beta) + \beta l$ with $\lambda(0)$ being both the length and the resting length at time
 112 zero and $F_c = 0$. With these choices and with a constant positive force F Eq. (1) has
 113 the following solution:

$$114 \quad l(t) = \frac{1}{(1-\beta)} \frac{F}{k} (1 - e^{-t(1-\beta)/\tau}) + \lambda(0) \quad (5)$$

115 Hence, the plasticity condition effectively reduces the body stiffness k to $k(1 - \beta)$
 116 enabling the system to reach a longer final size compared to the one allowed by the
 117 Kelvin-Voigt system alone, and increasing the relaxation time. By introducing

118 plasticity like in Eq. (4), the body progressively gains length at each stretching phase
119 (Fig S5d-d').

120

121 **1.3 The consequences of actin stability defects.** As reminded above, the intensity of
122 F_{epid} relies on two components: the constant contractility of the seam cell actomyosin
123 network, and the stiffness of the actin cables in DV cells. We can then represent it
124 like:

$$125 \quad F_{epid} = F_{seam} \alpha_{DV} \quad (6)$$

126 where the force F_{seam} represents the active force generated by myosin II in the
127 lateral cells and α_{DV} is the passive component given by the presence of actin
128 filament bundles in the DV cells. The biomechanical significance of equation (6) is
129 that both F_{seam} and α_{DV} positively contribute to F_{epid} , and that if one is absent $F_{epid}=0$.
130 This captures the fact that in the absence of myosin II there will be no pulling force
131 because the active component is absent, and that if actin cables are lost myosin II is
132 missing the resistance structure onto which it can pull, resulting in a null epidermis AP
133 force.

134 As shown in the main text (Fig. 3), the absence of SPC-1 and PAK-1, combined with
135 the mechanical input from muscles, induces actin integrity defects in DV cells. To
136 translate this situation in mathematical terms, we chose to write the passive
137 component α_{DV} as follows:

$$138 \quad \frac{d\alpha_{DV}}{dt} = \gamma \frac{dl}{dt} \quad \text{if} \quad \frac{dl}{dt} < 0 \quad \text{and} \quad \alpha_{DV} \geq 0 \quad (7)$$

139 where $\gamma \geq 0$ is a proportionality factor defined as a 'tearing factor'. The condition

140 $\frac{dl(t)}{dt} < 0$ means that α_{DV} decreases stepwise with time. In addition, we are imposing
 141 that $\gamma=0$ corresponds to an absence of tearing, like in wild-type embryos. The
 142 biological significance of this choice is the following: in phases of length decrease
 143 (net negative force), the overall circumference of the embryo should increase due
 144 to volume conservation (Fig. S4). In a background of unstable actin filaments (like in
 145 *spc-1 pak-1* double mutants) their resistance to stress originating from muscle activity
 146 would not be maintained. For this reason, α_{DV} should progressively decrease at each
 147 cycle. The condition $\alpha_{DV} \geq 0$ prevents α_{DV} from assuming negative values. Thus with
 148 α_{DV} decreasing, F_{epid} will progressively decrease, and as consequence the system
 149 length will shorten (see Fig. S5e and Fig. 4e).

150

151 **1.4 Equations summary.** In summary, we describe the embryo body as a plastic
 152 Kelvin-Voigt solid according to the following system of equations:

153 $\eta \frac{dl}{dt} = -k(l - \lambda) + F_{epid} + F_{muscles}$ (1) (Kelvin-Voigt with adjusting resting length

154 subject to $F_{epid} + F_{muscles}$)

155 $\frac{d\lambda}{dt} = \beta \frac{dl}{dt} H(\alpha_{DV})$ if $\frac{dl}{dt} > 0$, $0 \leq \beta < 1$ and $l - \lambda > F_c/k$ (5) (plasticity condition)

156 $\frac{d\alpha_{DV}}{dt} = \gamma \frac{dl}{dt}$ if $\frac{dl}{dt} < 0$ and $\alpha_{DV} \geq 0$ (7) (tearing condition)

157 With $F_{epid} = F_{seam} \alpha_{DV}$ as described in Eq.(4).

158 We assume that $\gamma=0$ for wild-type and *unc-112*, meaning that the resistance of dorso-
 159 ventral actin filament bundles remains unaffected by the body length changes
 160 caused by muscle activity.

161 The equations (1), (4), (5) and (7) have seven parameters: η , k , F_{seam} , $\alpha_{DV}(0)$, $F_{muscles}$, β
162 and γ . In order to reduce the parameter space we fixed some of them:

- 163 - for simplicity we set **$k=1$** ;
- 164 - from the laser ablation experiments performed in reference², the relaxation
165 time of epidermal actin filaments following the laser cut is in the order of a few
166 seconds. Being the relaxation time in a Kelvin-Voigt system given by $\tau = \eta/k$,
167 we set **$\eta=3$** so that the relaxation time is 3 seconds.
- 168 - F_{epid} is the multiplication of two parameters, and thus from the parameter
169 point of view can be considered as a single parameter that we formally set to
170 **$F_{seam}=1$** letting $\alpha_{DV}(0)$ as a free parameter.
- 171 - The size of the critical force F_c has been chosen to be half F_{seam} (**$F_c = 0.5$**).
172 Thereby, we consider that low intensity forces cannot trigger a plastic
173 response.
- 174 - Regarding $F_{muscles}$, we specified its details on the basis of the measured
175 contraction durations for embryos between 1.7 and 2-fold stages (Fig. S7). For
176 wild-type embryos, the duration of positive and negative periods has been set
177 to 6 seconds and the period of null contribution has been set to 15 seconds,
178 whereas for *spc-1* mutants and *spc-1 pak-1* double mutants the duration of
179 non-null activity has been set to 3 seconds and the duration of null
180 contribution is set to 15 seconds. In wild-type embryos, the intensity of $F_{muscles}$
181 has been left as a fit parameter together with α_{DV} , β and γ in order to be
182 determined by comparing with the data. The muscle force amplitude for *spc-*
183 *1* and *spc-1 pak-1* mutants has been set to **50% of the wt intensity** according
184 to our experimental observations.

185 To better adapt to the experimental observations, both F_{seam} and $F_{muscles}$ have been
186 modified by introducing an initial transient that sets their behavior from zero to the

187 regime of maximal intensity; the detailed form of these transients are reported in the
188 paragraph 'Refining model details'.

189

190 **1.5 Fitting procedure.** In order to determine the remaining four free parameters:
191 intensity of $F_{muscles}$, $\alpha_{DV}(0)$ (the initial value of actin strength), β (the plasticity factor)
192 and γ (tearing factor) we fitted different genotypes. We started with the muscle-
193 defective *unc-112* and wild-type embryos. As shown in Figs. 2 and S3, their
194 elongation rate is quite similar until the 2-fold stage, at which point *unc-112* embryos
195 have no muscle force ($F_{muscles}=0$), and they both have an identical actin pattern
196 ($\gamma=0$). Therefore, we fitted together the two elongation curves to find the values of
197 the three parameters $\alpha_{DV}(0)$, β and $F_{muscles}$ that capture the main features of the two
198 genotypes, and we refined the minor differences by allowing a 20% tolerance with
199 respect to the parameters values determined through the common fit. To estimate
200 the value of $F_{muscles}$ intensity in *spc-1* and *spc-1 pak-1* embryos, we considered that it
201 was half the wild-type value, based on the observation that the muscle
202 contraction/relaxation cycles were roughly twice shorter in these embryos (Fig. S7).
203 The notion that $F_{muscles}$ is lower in *spc-1* mutant embryos is consistent with the
204 observation that muscles make an angle of 20° with the anterior-posterior axis,
205 instead of 6° in wild-type embryos, predicting that their input should be reduced¹⁰.
206 The wild-type value of the plasticity factor was used as the upper limit for the
207 variability range of β for *spc-1* and *spc-1 pak-1* embryos. We then proceeded by
208 fitting the *spc-1* and *spc-1 pak-1* elongation curves to determine the values of
209 $\alpha_{DV}(0)$, β and the tearing factor γ (see below for the method).

210 The best fit parameters were determined by minimalizing the following cost function:

211
$$\xi = \sum_g \sum_i [(curve_{data}(i) - curve_{model}(i))^2]_g \quad (8)$$

212 where $curve_{data}(i)$ is the value of the data elongation curve at the time point i and
 213 $curve_{model}(i)$ is the solution of the model interpolated at the time point i ; g refers to
 214 the genotypes considered for a fit. To minimize the cost function, a Covariance Matrix
 215 Adaptation Evolution Strategy (CMA-ES) algorithm was used¹¹. The algorithm has an
 216 available Matlab code at http://www.lri.fr/~hansen/cmaes_inmatlab.html.

217 In the following table we report the values of the fitted parameters for the different
 218 genotypes presented in the Fig. 4j and S5f. Errors have been obtained by fitting the
 219 mean curves plus and minus their standard errors respectively and by taking the
 220 maximal deviation from the parameters obtained from the mean.

221

	wild-type	<i>unc-112</i>	<i>spc-1</i>	<i>spc-1 pak-1</i>
Initial actin stiffness $\alpha_{DV}(0)$	1.06 ± 0.05	1.13 ± 0.01	0.84 ± 0.03	0.76 ± 0.03
Plasticity factor β	0.12 ± 0.06	0.05 ± 0.01	0.06 ± 0.02	0.02 ± 0.03
Intensity $F_{muscles}$	5.0 ± 2.5	∅	2.5	2.5
Tearing factor γ	∅	∅	0.04 ± 0.01	0.12 ± 0.01

222

223 **1.6 Refining model details.** Ablation experiments in seam cells² have shown that the
 224 circumferential stress is not constant during development but increases from the 1.3-
 225 fold to 1.7-fold stages. To better adapt our model to this observation, we introduced
 226 a time dependent function for F_{epid} that saturates to a plateau value within a time τ_s :

227
$$F_{epid}(t) = F_{seam} \alpha_{DV} (1 - \exp(-\frac{t}{\tau_s}))^{h_s} \quad (9)$$

228 where h_s is an exponent that sets the steepness of the function in reaching the
 229 plateau and $F_{seam} \alpha_{DV}$ is the plateau value. We set $\tau_s=50$ min and $h_s=4$, since this
 230 choice produces slow elongation for the first 50 minutes, as was observed for all the
 231 genotypes (see Fig. 2). Much like the epidermis force, the muscle force $F_{muscles}$ also
 232 starts at a small amplitude, then progressively evolves with a behavior similar to
 233 equation (8):

234

$$235 \quad F_{muscles}(t) = \begin{cases} A \theta(t) \left(1 - \exp\left(-\frac{(t-t_{init})}{\tau_m}\right)\right)^{h_m} & \text{for } t > t_0 \\ 0 & \text{otherwise} \end{cases} \quad (10)$$

236

237 where A is the maximum amplitude, $\theta(t)$ is the periodic function made of a
 238 composition of cosines described in Fig. S5c; h_m sets the steepness of the function to
 239 reach the plateau; t_0 is the time at which muscles get active and τ_m is the time
 240 necessary for the function to reach its plateau.

241 We started to measure the elongation curves immediately before the comma stage,
 242 which represents our initial time. We set $t_{init}=90$ min, since around this point of
 243 elongation wild type embryos are 1.7-fold long and they start to contract muscles;
 244 we also assume that after the 2-fold stage the muscle force has reached its
 245 maximum amplitude, which is why we fixed $\tau_m=15$ min (the time necessary for a wild
 246 type embryo to go from 1.7 to 2 fold; h_m has been arbitrarily set to 1).

247 The model summarized in Eqs. (5) and (9) has no plateau; however *C. elegans*
 248 embryos extend up to 4-fold within ≤ 150 minutes, then stay at this length for ~ 100
 249 additional minutes before hatching. We do not know why the body length stops at 4-

250 fold and this issue is not the focus of this work. We can speculate that the ability of
 251 the body to remodel is not illimited and may also be restricted by cuticle secretion. In
 252 this framework, we can modify Eq.(5) by introducing a multiplicative term under the
 253 form of a Hill function to account for the saturation of the elongation:

$$254 \quad \frac{d\lambda}{dt} = \beta \frac{dl}{dt} H(\alpha_{DV}) Hill(\lambda) \quad \text{if } \frac{dl}{dt} > 0, 0 \leq \beta < 1 \text{ and } l - \lambda > F_c/k \quad (11)$$

255 where $Hill(\lambda) = \frac{L^d}{L^d + \lambda^d}$, with L upper threshold for the resting length and d an
 256 exponent¹². We set **L=3.2** and **d=15** to account for a rapid saturation of elongation
 257 after the 3.2-fold stage, since the wild type embryos show, after this length, a rapid
 258 reduction of the elongation rate. By introducing these additional functions, we can
 259 improve the agreement between the data and the model results (Fig. 4j). The
 260 Equations presented above have been solved numerically by the Euler method
 261 implemented in a Matlab script, with the choice of initial length $l(0) = \lambda(0) = 1$.

262 Below is a summary of the parameter values that we fixed on the basis of
 263 experimental observations and therefore were not used as free parameters.

264

Spring	$k=1$
Dashpot	$\eta=3$
F_{epid}	$F_{seam}=1, \tau_s=50, h_s=4$
$F_{muscles}$	$\tau_m=15, h_m=1, t_{init}=90$
Saturation function	$L=3.2, d=15$
Critical force F_c	0.5

265
 266

267 **1.7 Predictiveness of the model.** Here we test the model described above by
268 comparing the predicted elongation curves with those observed with another
269 genotype.

270 When muscle activity was compromised in *spc-1 pak-1* embryos, the retraction
271 phenotype was prevented (Fig. 2). From equations (1) and (7), if $F_{muscles}$ equals zero,
272 then there should be no retraction. Moreover, by fitting the *unc-112; spc-1 pak-1*
273 elongation curve we obtained a value of $\alpha_{DV} = 0.64 \pm 0.04$.

274

275 **1.8 Kelvin-Voigt-type model or Maxwell-type model?** In the present work we
276 decided to model the *C. elegans* embryo as a visco-elastic solid with the ability to
277 rearrange its resting length during elongation, and we show that this description can
278 account both for elongation and retraction by changing the behavior of the
279 epidermal force. The Maxwell model⁷, as a dashpot and a spring in series, would
280 also account for elongation when the system is under the effect of a positive force.
281 However, a Maxwell-type material keeps extending as soon as a positive force acts
282 on it and this behavior would not account for the muscle-defective *unc-112*
283 embryos. Indeed, these embryos elongate till about 2-fold under the effect of the
284 epidermal force, but does not grow further in the absence of muscle activity,
285 whereas a Maxwell model would predict that its elongation continues.

286

287 **2- Methods**

288 **Animal strains, conditions of maintenance**

289 The list of strains used is presented in Supplementary Table S4. Animals were
290 propagated on NGM agar plates as described previously¹³. Animals were held at a
291 standard 20°C temperature except for experiments implicating *sma-1(ru18); pak-*
292 *1(tm403)*, in which case L4 animals were shifted to 15°C before egg laying and
293 measures were performed on their offsprings.

294

295 **Yeast Two Hybrid Screening**

296 Yeast two-hybrid screening was performed by Hybrygenics Services (Paris, France).
297 The bait component was the N-terminal 294 amino acids of PAK-1 (including the
298 regulatory region of the protein) fused as a C-terminal fusion to LexA. The construct
299 was used as a bait to screen at saturation a highly complex, random-primed
300 *C.elegans embryo* cDNA library¹⁴. Screening involved a mating approach on a
301 medium lacking Trp, Leu and His, supplemented with 0.5 mM 3-aminotriazole¹⁵. The
302 strongest hits of the screen can be found in Supplementary Table 1. Reciprocal
303 screens using the spectrin repeat #9 or the SH3 domains of SPC-1 as baits identified
304 PAK-1 region 160-206, and no other meaningful prey in the context of the present
305 study.

306

307 **RNAi enhancer screen and RNAi assays**

308 Two RNAi screens were performed in the *pak-1(tm403)* and in the *spc-1(ra409)*
309 backgrounds along with a wild type control. A collection of 356 essential genes from
310 the Ahringer RNAi library¹⁶, including adhesion proteins, signaling proteins,
311 phosphatases, kinases, cytoskeleton-associated proteins and proteins important in

312 epithelial morphogenesis, was assembled (Table S1), based on a previous screen
313 performed in the *git-1(tm1962)* background (C. Gally and M. Labouesse,
314 unpublished). The screen was performed in liquid culture on 96-well plates and RNAi
315 knockdown was induced by feeding as described¹⁷. The primary screen was based
316 on enhanced lethality and body morphology defects; the secondary screen
317 focused on very short larvae and elongation defects. We took DIC images for body
318 length measurements and performed DIC timelapse imaging for the strongest
319 candidates. For specific genes, RNAi was always induced by injection after
320 preparing the double-stranded RNA (dsRNA) with the Ambion mMessage
321 mMACHINE[®] kit and purifying the dsRNA with the Qiagen RNeasy[®] MinElute[®] Cleanup
322 kit¹⁸. The embryos were analyzed from 24h to 48h post-injection.

323

324 **Fluorescent translational reporter constructs**

325 A 12633 bp genomic sequence including the *spc-1* coding sequence and a 3 kb
326 promoter was inserted in frame with the GFP coding sequence present in the
327 pPD95.75 vector (Addgene, Cambridge USA). To create the *Ppak-1::pak-1::mKate*
328 reporter construct, first an mKate-containing backbone was created by exchanging
329 the GFP-coding sequence of the pML1572, *Plin-26::vab-10(ABD)::GFP* plasmid (Gally
330 et al., 2009). In a following cloning step a 8204 bp genomic sequence, including the
331 *pak-1* coding sequence and a 4.5 kb promoter was inserted in frame with the mKate
332 coding sequence present in the vector.

333 To test if SPC-1::GFP could rescue the function of SPC-1, we first crossed the different
334 transgenic animals of a wild type background with *mnDp33; spc-1(ra409)* animals
335 (strain DM3409), and F1 transgenic males again with DM3409 to establish *mnDp33;*
336 *spc-1(ra409); Ex[spc-1::GFP]*. Rescuing transgenes were recognized because all

337 viable progeny was GFP-positive and all (or most) non-viable progeny was GFP-
338 negative, reflecting the loss of the *mnDp33* balancer. To attempt *mnDp33*
339 segregation, we repeatedly transferred single GFP-positive mothers over four
340 generations and examined their progeny, starting from at least two independent
341 extrachromosomal arrays per construct. Thereby, we successfully obtained viable
342 *spc-1(ra409); Ex[P_{spc-1}::*spc-1(+)*::*gfp*]* animals, and then viable *spc-1(ra409) pak-*
343 *1(tm403); Ex[P_{spc-1}::*spc-1(+)*::*gfp*]* animals through crossing and meiotic
344 recombination, which segregated very short retracted GFP-negative embryos. For
345 the FHOD-1 constitutively active construct, a 8283 bp genomic sequence of *fhod-1*
346 (gift from David Pruyne¹⁹) deleted for part of the FH2 domain and the DAD domain
347 was inserted under the control of the epidermis-specific 432 bp *dpy-7* promoter.
348 Deletion of the DAD alone was obtained by inserting back the FH2 domain in the
349 FH2/DAD deleted construct using Hifi DNA assembly cloning kit (New England
350 Biolabs); the FH1/FH2 deleted construct was obtained by deleting the FH1 domain
351 using the Q5 site directed mutagenesis kit protocol (New England Biolabs). All cloning
352 steps relied on the use of the Phusion High-Fidelity DNA Polymerase reaction kit (Fisher
353 Thermo-Scientific); the constructs were subsequently verified by sequencing. The
354 constructs were injected at 10 ng/μl plasmid construct, with 150 ng/μl pBSK + 5 ng/μl
355 pCFJ90 (*P_{myo-2}::mcherry*) as co-injection markers for the *spc-1::gfp* constructs, or 100
356 ng/μl pRF4 (*rol*) co-injection marker for *pak-1::mKate*.

357

358 **Fluorescence imaging**

359 DIC images for time-lapse videos were obtained using a Leica DM6000 microscope
360 with a Leica LAS-AF software, using a Leica DMRXA2 upright microscope equipped
361 with a Peltier platform. Observations were done under a 40X oil immersion objective.

362 Mothers were cut up to gain early-staged embryos, which were then transferred
363 onto thin 5% soft agarose pads in a drop of M9. Z stack image series with a 1,5 μm Z
364 step distance were taken in every 5 minutes during 6 hours. ImageJ software was
365 used to quantify the embryonic length from the end of ventral enclosure/onset of
366 elongation, by taking a "segmented line" through the midline of the embryos from
367 head to tail. To image the coupling between actin bundles displacement in the
368 epidermis and muscle contractions, we used a double reporter strain carrying the
369 epidermal *Pdpy-7::LifeAct::GFP* and muscle *Pmyo-3::his-24::mCherry* transgenes
370 (ML2113, see Table S4), and a spinning-disk DMI6000 Leica microscope equipped
371 with an Andor software (experiments for Fig. 1). Series of five Z planes (1 epidermal +
372 4 muscle) were imaged continuously for 5 min, with 0,5 μm Z steps and no averaging.
373 The time interval between two Z series was 360 ms. The measurement of actin
374 displacement was done according to the same strategy, and was done using a
375 CSUX1-A1 spinning-disk mounted on a Zeiss Axio Observer Z1 inverted microscope
376 with a Roper Evolve camera controlled by the Metamorph software, and a 100x oil
377 immersion objective (experiments for Fig. S9). A Z-stack of 4 focal planes with 0,5 μm
378 step size was acquired using a streaming acquisition mode. The time between two
379 acquisitions was 0.41 second during 300 time frames. To synchronize embryos,
380 mothers were put on an empty NGM agar plate to lay eggs for a short time window,
381 and embryos were left to develop until the stage of interest. (For the analyse of the
382 contraction see next section on **Image analysis**. To analyse the *in vivo* co-localisation
383 between *PAK-1::mKate*, *ABD::mcherry* and *SPC-1::GFP*, we used the Zeiss/Roper
384 spinning disk microscope under a 100X oil immersion objective, keeping the laser
385 intensity at a constant level throughout the experiments. Image processing and
386 computing the co-localisation coefficient was done using the Volocity software.

387

388 **Image analysis and quantification of actin filament contraction, continuity and**
389 **orientation**

390 The analysis of mechanical displacement in the epidermis was performed on the
391 movies of the dorso-ventral actin layer by measuring the distance over time
392 between two landmarks across the region of contraction. The landmarks were set
393 manually on a frame showing relaxed tissue and tracked all over the contraction till
394 the subsequent relaxed state. Landmarks tracking was performed using a statistical
395 template matching approach²⁰. The method was implemented as an ImageJ plugin
396 (<http://sites.imagej.net/Julienpontabry/>), giving as output the landmarks locations,
397 their distance across time and the Kymographs. The curves show a pre-contraction
398 state, a minimum (the maximal contraction point) and an ending part where the
399 distance progressively increases again (see Fig. 1B'' and Fig. S7). After a smoothing
400 and interpolation of the curves, the starting, ending and maximal contraction points
401 were extracted by studying the time derivative of the distance and by setting a
402 threshold on the distance itself. Finally the contraction time was computed as the
403 difference between the ending time and the initial time. The analysis of the curves
404 and statistics were done using a MatLab script. All images were analysed using the
405 ImageJ (Fiji) software (NIH, Bethesda, Maryland, USA; <http://rsb.info.nih.gov/ij/>) and
406 MATLAB R2014b (The MathWorks Inc., Natick, MA). To study the features of actin
407 pattern, we imaged embryos that were put to sleep by oxygen deprivation through
408 a high concentration of bacteria with the Zeiss/Roper spinning-disk using a 100X oil
409 immersion objective. For each experiments, a Z-stack of 16 focal planes with 0,2 µm
410 step size was acquired. On the original maximum z projection created by imageJ, a
411 manual ROI was defined on the dorso-ventral cells (Fig. 3) from which a high pass
412 filter in the Fourier space was applied to select only structures smaller than 10 pixel of
413 diameter (Fig. 3).

414

415 **Continuity:** The filtered ROIs were binarized by setting to one all the pixels with a
416 value bigger than zero and setting to zero all the other pixels. The resulting structures
417 were then fitted by ellipses from which the length of the Major axis was extracted as
418 a measure of the length of the actin filament. The longer actin filaments are those
419 presenting a more uniform fluorescence along their length (showing higher
420 continuity). By contrast, short segments result from discontinuity in the fluorescence
421 signal. To avoid noise only segments longer than 4 pixels have been considered for
422 the analysis.

423

424 **Anisotropy of the orientation:** The same filtered ROI used for continuity measurements
425 were used to measure the distribution of cable orientation. Fast Fourier Transform
426 (FFT) of these ROIs was computed in order to work in the frequency domain and
427 more easily identify repetitive patterns. The resulting power spectrum of the ROIs was
428 represented in polar coordinates in order to extract the distribution of angles of ROI
429 pattern²¹. The method was implemented in an ImageJ plugin
430 (<http://sites.imagej.net/Julienpontabry/>), giving as output the angular distribution. In
431 order to compare the distributions coming from different images the distributions
432 were normalized by their integral (Fig. 3). The more the pattern consists in structures
433 oriented in a preferred direction (the more anisotropic), the highest is the peak of the
434 distribution in that direction. In the case of an isotropic pattern, the angular
435 distribution should show a flat behavior. As an estimate of the pattern anisotropy, the
436 prominence of the highest peak of the angular distribution was considered (Fig. 3
437 and Fig. S4). The analysis of the angular distributions and statistics was performed by
438 a MatLab script. All images were analysed using the ImageJ (FiJI) software (NIH,
439 Bethesda, Maryland, USA; <http://rsb.info.nih.gov/ij/>) and MATLAB R2014b (The

440 MathWorks Inc., Natick, MA). All matlab scripts used for the present analysis are
441 available upon request.

442

443 **Statistical Analysis**

444 For elongation curves, standard deviation was measured. For L1 length
445 measurement and rescue experiments we perform unpaired t-test and ANOVA test
446 using GraphPad Prism 5.00 (San Diego, California, USA) and Excel. For contraction
447 time, actin continuity and orientation, we applied for all genotypes a paired t-test
448 using Matlab.

449

450

451 **3-Supplementary figures**

452 **Supplementary Figure 1:** Local bending of epidermal actin filaments due to muscle
453 contractions

454 **(a-c)** Displacement of actin filament marked by LifeAct::GFP at three time points in
455 wild-type embryos. Note their local bending (yellow box). Scale bar, 10 μ m.

456

457 **Supplementary Figure 2:** Genes required to maintain embryonic elongation.

458 **(a-g)** The proteins GIT-1 and PIX-1 are acting upstream of PAK-1 in the mechano-
459 transduction pathway promoted by muscle contractions, and their loss in the
460 absence of *spc-1* also triggers a retraction phenotype. **(a)** Elongation curves and **(b-**

461 **g)** terminal phenotypes of **(b)** *pak-1(tm403)*; **(c)** *git-1(tm1962)*; **(d)** *pix-1(gk416)*; **(e)**
462 *spc-1(RNAi) pak-1(tm403)*; **(f)** *git-1(tm1962) spc-1(RNAi)* **(g)** *spc-1(RNAi) pix-1(tm416)*.

463 **(h-k)** The *pak-1(tm403)* mutation also showed synergistic body morphology defects
464 when combined with the null allele *sma-1(ru18)* in the apical β -spectrin SMA-1, but

465 not as pronounced as for loss of the α -spectrin SPC-1. Quantification of the length at
466 hatching **(h)** and terminal phenotypes of **(i)** *pak-1(tm403)*, **(j)** *sma(ru18)*, **(k)**

467 *sma(ru18); pak-1(tm403)* L1 hatchlings. **(l)** Elongation curves and **(m-n)** terminal
468 phenotypes of **(m)** *unc-112(RNAi)* (n=5) and *unc-112(RNAi); pak-1(tm403)* (n=8)

469 embryos. **(o)** Terminal phenotype of *unc-112(RNAi); spc-1(ra409)* obtained by
470 inducing *unc-112(RNAi)* in the strain ML2436 bearing a rescuing extrachromosomal

471 *spc-1::gfp* array and looking for embryos having lost the array; due to high stability of
472 the array few embryos were obtained despite numerous repeats (n=4), which all had

473 the phenotype illustrated here and is similar to that of *spc-1(ra409)* alone. Scale bars:

474 17 μ m **(b-g; m-o)**; 50 μ m **(i- k)**.

475

476 **Supplementary Figure 3: PAK-1 and SPC-1 colocalized with actin filaments**

477 **(a)** Distribution of PAK-1::mKate in a late embryo (n=20). Enlarged pictures of PAK-1
478 showing a filamentous distribution in the dorso-ventral epidermis similar to actin
479 filaments. Scale bar: 25 μ m. **(b)** Fluorescence pictures of PAK-1::mKate (red) and SPC-
480 1::GFP (green): the panel shows the colocalization image for the most apical focal
481 planes (top image), and full XZ (green panel) and YZ (red panel) projections. The
482 level of co-localization is high based on Pearson's correlation coefficient (0.7-0.9,
483 n=20). The highest level of co-localization is detected at the apical cortex. **(c)**
484 Fluorescence pictures of *Plin-26*::VAB-10(ABD)::mCherry (red) and SPC-1::GFP (green):
485 the panel shows the colocalization image for the most apical focal planes (top
486 image), and full XZ (green panel) and YZ (red panel) projections. The level of co-
487 localization is high based on Pearson's correlation coefficient (0.7-0.9, n=8). The co-
488 localization is almost exclusively detected at the apical cortex. The gene *lin-26* drives
489 expression in the epidermis; VAB-10(ABD) corresponds to the two actin-binding
490 domains (calponin homology) of the protein VAB-10. Scale bar, 10 μ m.

491

492 **Supplementary Figure 4: Actin filament continuity and orientation at three elongation**
493 stages

494 Actin filaments were visualised using a *Pepid*::*Lifeact*::*GFP* construct and
495 characterised as outlined in Fig. 3a (the *Pepid* promoter corresponds to *Pdpy-7*). **(a)**
496 Actin filament continuity. The graph represents the length (in pixels) along the
497 circumferential axis of actin filaments in early, mid and late (corresponding to 1.7-
498 fold, 2-fold and 3-fold equivalent stages in a wild-type embryo, respectively)
499 embryos of wild-type (early n=12, mid n=19, late n=16), *pak-1(tm403)* (early n=16, mid
500 n=21, late n=16), *spc-1(RNAi)* (early n=15, mid n=21, late n=20), and *spc-1(RNAi) pak-*
501 *1(tm403)* (early n=12, mid n=17, late n=26) genotypes. **(b)** Actin filament orientation

502 based on Fast Fourier Transform and binarisation. Wild type (early n=12, mid n=18,
503 late n=14), *pak-1(tm403)* (early n=16, mid n=20, late n=16), *spc-1(RNAi)*(early n=14,
504 mid n=18, late n=18), and *pak-1(tm403) + spc-1(RNAi)* (early n=12, mid n=18, late
505 n=21). Note that the characteristics of actin filaments in *spc-1(RNAi) pak-1(tm403)*
506 embryos differ mostly at the equivalent of the two-fold stage when muscles become
507 active. At earlier and later stages, *spc-1(RNAi)* embryos and *spc-1(RNAi) pak-*
508 *1(tm403)* embryos become similar. **(c-f)** Epidermal actin filaments visualized with the
509 *Pepid::Lifeact::GFP* construct in wild-type (C), *spc-1(RNAi) pak-1(tm403)* (D) embryos
510 (panels are from Fig. 3). * $<0,05$; ** $<0,001$; *** $<0,0001$; ns, not significant.

511

512 **Supplementary Figure 5:** Change in embryo diameter during elongation

513 **(a-b'')** Fluorescence micrographs of embryos expressing the *Pepid::Lifeact::GFP*
514 construct in the epidermis at three elongation stages early, middle and late
515 (corresponding to 1.7-fold, 2-fold and 3-fold equivalent stages in a wild-type embryo,
516 respectively) for wild-type **(a-a'')** and *spc-1(RNAi) pak-1(tm403)* embryos **(b-b'')**; the
517 *Pepid* promoter corresponds to *Pdpy-7*. The yellow lines correspond to the segments
518 used to measure the dorso-ventral width of the V1 seam cell. **(c)** Quantification of
519 the average V1 cell width normalized to the initial width during elongation in four
520 genotypes. **(d)** Quantification of the average dorso-ventral width at the level of the
521 V1 seam cell, which was calculated using the measured embryo length and V1 cell
522 width, taking into consideration the conservation of the total embryo volume. Each
523 point in panel **(c)** and **(d)** represents between 7 and 21 embryos; error bars are
524 standard errors. A notable feature of *spc-1(RNAi) pak-1(tm403)* embryos is that the
525 circumferential dimension of the seam cells decreased much more than that of their
526 DV cell, which most likely reflects the actin filament integrity defects combined with
527 a F_{seam} force largely unchanged.

528

529 **Supplementary Figure 6:** Time-dependent length of a Kelvin-Voigt model in different
530 conditions.

531 **(a-a')** A generic Kelvin-Voigt system subject to a constant force F_{epid} and its
532 predicted elongation change using $F_{epid}=0.85$ (A'). **(b-b')** A similar system subject to
533 two forces, F_{epid} and an oscillating force $F_{muscles}$ with the properties depicted in (E),
534 and predicted elongation change using $F_{epid}=0.85$ (B'). As the pulsatile force induces
535 both compression and stretching (see Fig. S6), its net input on elongation is transient
536 and the system oscillates around the maximal value reached without $F_{muscles}$. **(d-d')** A
537 Kelvin-Voigt system with mechanical plasticity introduced according to Eqs. (1,4-7),
538 and predicted elongation change using $F_{epid}=0.85$, $\beta=0.10$ and $F_c=0$ according to
539 Eqs. (1,4-7). **(e)** A Kelvin-Voigt system in which the plasticity is defective ($\beta=0$), and
540 there is actin tearing according to Eq. (7) inducing a progressive reduction of F_{epid} ,
541 and predicted elongation change using $F_{epid}=0.85$, with an initial value of the passive
542 component $\alpha_{DV}(t=0)=1$ and the tearing factor $\gamma=0.15$ (D'). **(e' Inset)** Behavior of
543 $\alpha_{DV}(t)$. **(c)** In all conditions $F_{muscles}$ is a periodic function with positive and negative
544 steps of duration of 6 seconds alternating with periods of null value of duration 15
545 seconds modulated by a cosine function. In **(a-e')** the elastic constant of the spring is
546 $k=1$, the initial resting length has the value $\lambda(t=0)=1$, and the viscosity value is $\eta=10$. **(f)**
547 Result of the fit for the following genotypes: *wt*, *spc-1 pak-1*, *unc-112* and *spc-1*
548 according to Eqs. (1,4-7). The values of the parameters are specified in paragraphs
549 1.4, 1.5 and 1.6.

550

551 **Supplementary Figure 7:** Comparable retraction phenotypes after the combined loss
552 of SPC-1 and PAK-1 or SPC-1 and FHOD-1

553 **(a)** Elongation curves of the single and double deficient embryos indicated on the
554 left, **(b-h)** and corresponding terminal phenotypes at hatching: wild-type (n=5),
555 *fhod-1(tm2363)* (n=6) *fhod-1(RNAi)* (n=7) *pak-1(tm403)* (n=5) *fhod-1(RNAi);*
556 *pak1(tm403)* (n=7), *spc-1(RNAi) pak-1(tm403)* (n=8), *fhod-1(tm2363);spc-1(RNAi)*
557 (n=9). Scale bar, 25 μ m.

558

559 **Supplementary Figure 8:** PAK-1 and FHOD-1 form aggregates in *spc-1(RNAi)* loss of
560 function.

561 **(a-a')** PAK-1::GFP localisation in wild-type and *spc-1(RNAi)* embryos. Yellow box, area
562 enlarged below the panel. Note the punctae in SPC-1 deficient embryos. **(b-b')**
563 FHOD-1 localization in wild-type and *spc-1(RNAi)* embryos. Note the aggregates
564 (arrowheads). Note also that FHOD-1::GFP displayed a filamentous organization
565 reminiscent of actin filaments. Scale bar: 10 μ m.

566

567 **Supplementary Figure 9:** Actin displacement ratio

568 **(a-d)** Individual displacement tracks of actin filaments visualized with a *Pdpy-*
569 *7::Lifeact::GFP* marker specifically expressed in the epidermis of wild type **(a)**, *pak-*
570 *1(tm403)* **(b)**, *spc-1(RNAi)* **(c)** and *spc-1(RNAi) pak-1(tm403)* **(d)** embryos at a stage
571 equivalent to 2-fold in a wild-type embryo. Scale bar: 10 μ m. **(e)** Typical kymographs
572 of *Pepid::Lifeact::GFP*-labeled actin filaments in wild-type and *spc-1(RNAi) pak-*
573 *1(tm403)* embryos from which the tracks in a-d were derived. Yellow dots correspond
574 to landmarks for quantitative analysis. **(f)** Quantification of the displacement duration
575 in (N=embryo/ n=contraction): wild-type, N=11/n=51; *pak-1(tm403)*, N=11/n=26; *spc-*
576 *1(RNAi)*, N=11/n=73; *spc-1(RNAi) pak-1(tm403)*, N=11/n=89. *<0,05; **<0,001;
577 ***<0,0001; ns, not significant.

578

579 **4-Caption for movies**

580 **Movie 1:** Combined DIC timelapse movie. Image acquisition was every 5 minutes in
581 wild-type, *pak-1(tm403)*, *spc-1(RNAi)*, *spc-1(RNAi) pak-1(tm403)* embryos. Scale Bar,
582 10 μ m.

583 **Movie 2:** Combined DIC timelapse movie of *unc-112(RNAi)* and *unc 112(RNAi); spc-*
584 *1(RNAi) pak-1(tm403)* embryos. Scale Bar, 10 μ m.

585 **Movie 3:** Combined DIC timelapse movies of *spc-1(ra409)* and *fhod-1(RNAi); spc-*
586 *1(ra409)* embryos. Scale Bar, 10 μ m.

587 **Movie 4:** Fluorescence movie showing the displacement of actin filaments labelled
588 with *Pdpy-7::lifeact::GFP* in the epidermis. Time acquisition is 0.41 s in wild-type, *pak-*
589 *1(tm403)*, *spc-1(RNAi)* and *spc-1(RNAi) pak-1(tm403)* embryos. Scale Bar, 10 μ m.

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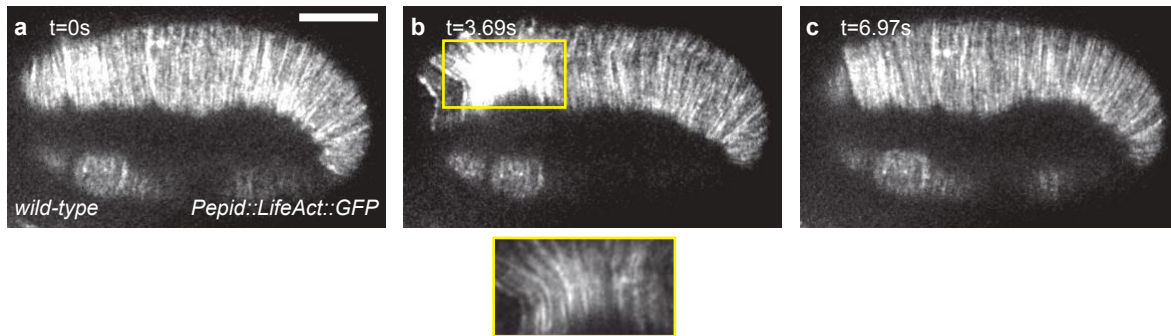
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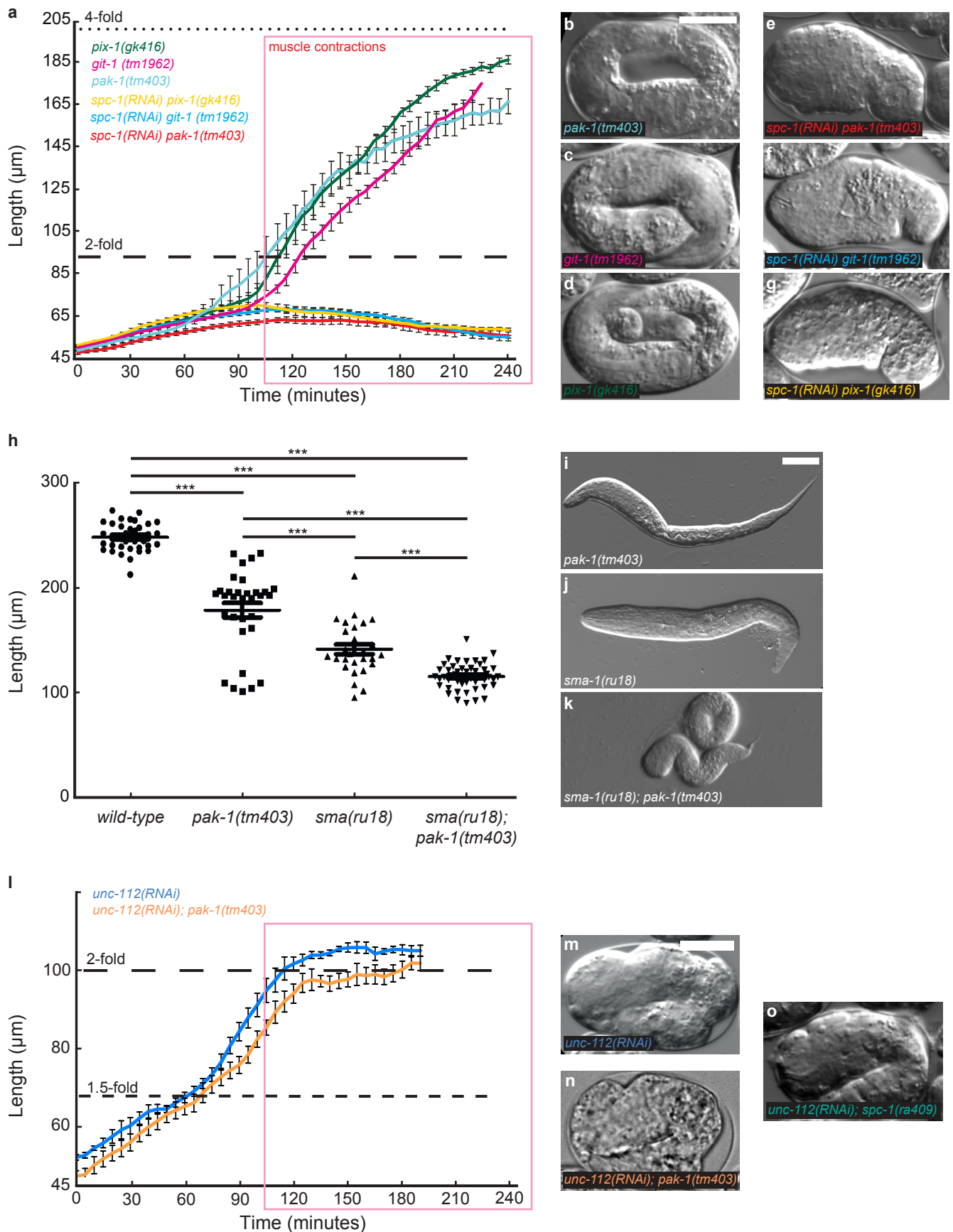
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Supplementary Figure 1: Local bending of epidermal actin filaments due to muscle contractions

(a-c) Displacement of actin filament marked by LifeAct::GFP at three time points in wild-type embryos. Note their local bending (yellow box). Scale bar, 10 μ m.



Supplementary Figure 2: Genes required to maintain embryonic elongation.

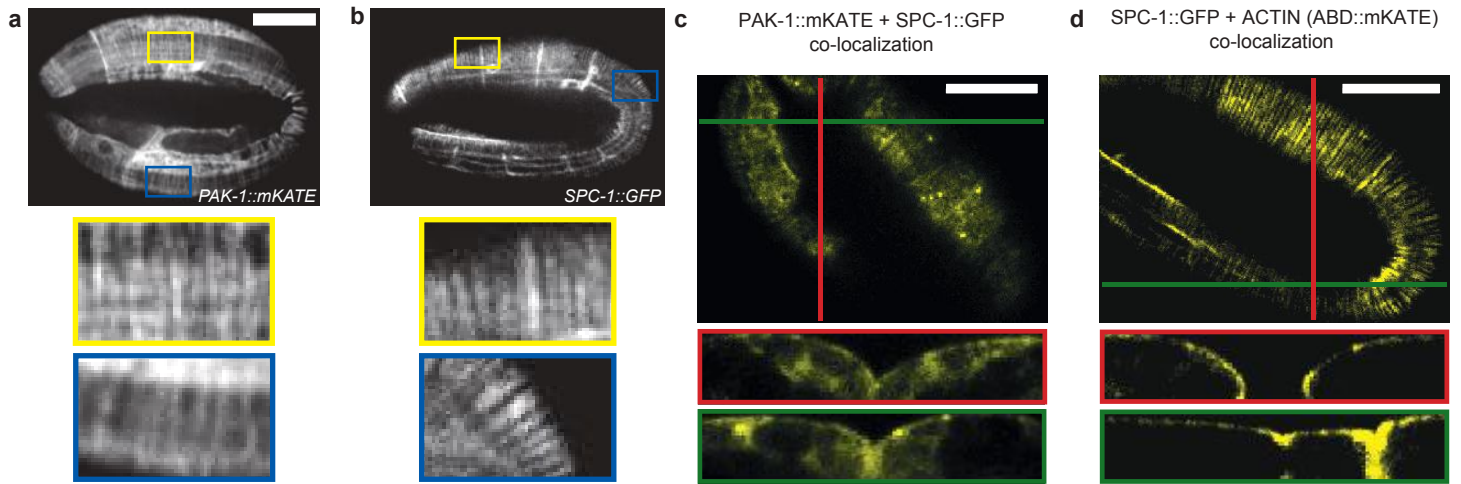
(a-g) The proteins GIT-1 and PIX-1 are acting upstream of PAK-1 in the mechano-transduction pathway promoted by muscle contractions, and their loss in the absence of *spc-1* also triggers a retraction phenotype.

(a) Elongation curves and (b-g) terminal phenotypes of (b) *pak-1(tm403)*; (c) *git-1(tm1962)*; (d) *pix-1(gk416)*; (e) *spc-1(RNAi) pak-1(tm403)*; (f) *git-1(tm1962) spc-1(RNAi)* (g) *spc-1(RNAi) pix-1(tm416)*.

(h-k) The *pak-1(tm403)* mutation also showed synergistic body morphology defects when combined with the null allele *sma-1(ru18)* in the apical β -spectrin SMA-1, but not as pronounced as for loss of the α -spectrin SPC-1. Quantification of the length at hatching (h) and terminal phenotypes of (i) *pak-1(tm403)*, (j) *sma(ru18)*, (k) *sma(ru18); pak-1(tm403)* L1 hatchlings.

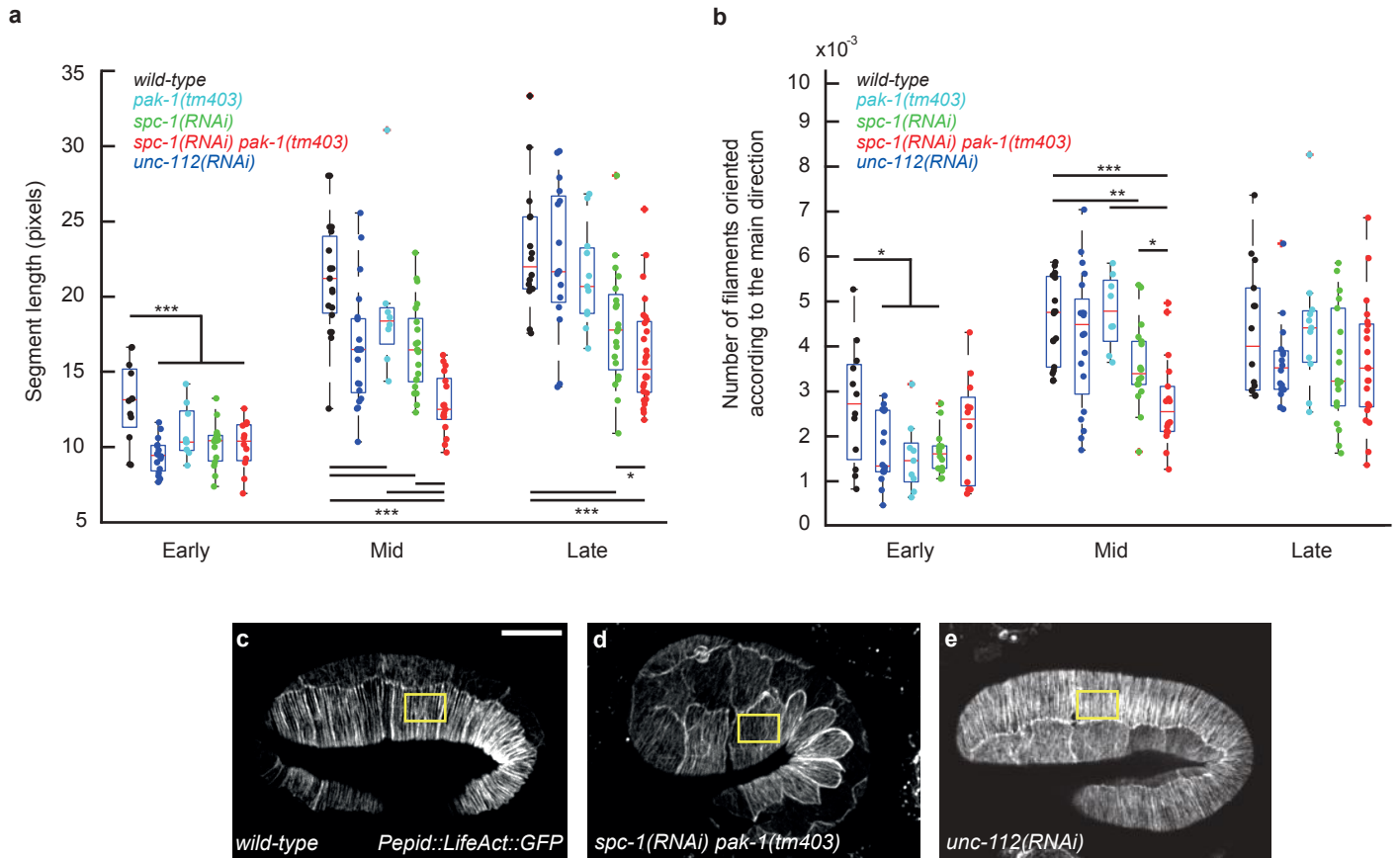
(l) Elongation curves and (m-n) terminal phenotypes of (m) *unc-112(RNAi)* (n=5) and *unc-112(RNAi); pak-1(tm403)* (n=8) embryos.

(o) Terminal phenotype of *unc-112(RNAi); spc-1(ra409)* obtained by inducing *unc-112(RNAi)* in the strain ML2436 bearing a rescuing extrachromosomal *spc-1::gfp* array and looking for embryos having lost the array; due to high stability of the array few embryos were obtained despite numerous repeats (n=4), which all had the phenotype illustrated here and is similar to that of *spc-1(ra409)* alone. Scale bars: 17 μm (b-g; m-o); 50 μm (i-k).



Supplementary Figure 3: PAK-1 and SPC-1 colocalized with actin filaments

- (a) Distribution of PAK-1::mKate in a late embryo (n=20). Enlarged pictures of PAK-1 showing a filamentous distribution in the dorso-ventral epidermis similar to actin filaments. Scale bar: 25 μ m.
- (b) Fluorescence pictures of PAK-1::mKate (red) and SPC-1::GFP (green): the panel shows the colocalization image for the most apical focal planes (top image), and full XZ (green panel) and YZ (red panel) projections. The level of co-localization is high based on Pearson's correlation coefficient (0.7-0.9, n=20). The highest level of co-localization is detected at the apical cortex.
- (c) Fluorescence pictures of Plin-26::VAB-10(ABD)::mCherry (red) and SPC-1::GFP (green): the panel shows the colocalization image for the most apical focal planes (top image), and full XZ (green panel) and YZ (red panel) projections. The level of co-localization is high based on Pearson's correlation coefficient (0.7-0.9, n=8). The co-localization is almost exclusively detected at the apical cortex. The gene *lin-26* drives expression in the epidermis; VAB-10(ABD) corresponds to the two actin-binding domains (calponin homology) of the protein VAB-10. Scale bar, 10 μ m.



Supplementary Figure 4: Actin filament continuity and orientation at three elongation stages

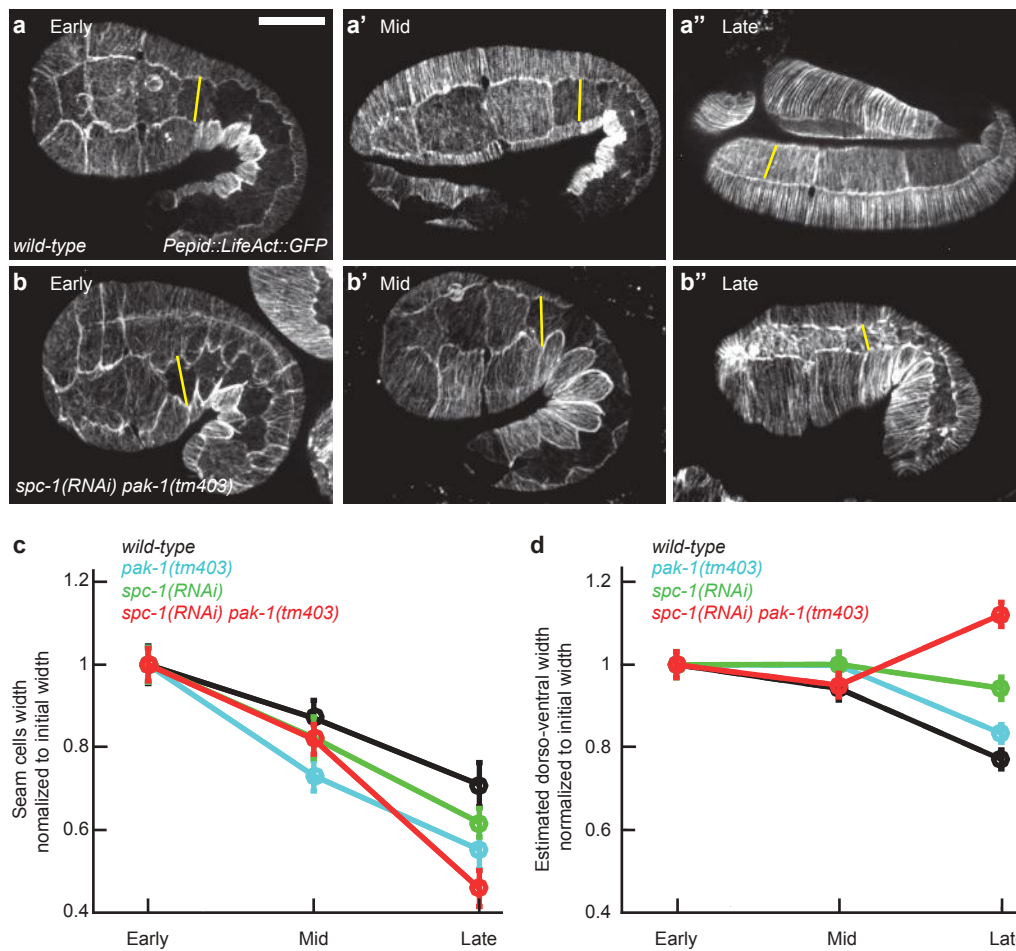
Actin filaments were visualised using a *Pepid::Lifeact::GFP* construct and characterised as outlined in Fig. 3a (the *Pepid* promoter corresponds to *Pdpy-7*).

(a) Actin filament continuity. The graph represents the length (in pixels) along the circumferential axis of actin filaments in early, mid and late (corresponding to 1.7-fold, 2-fold and 3-fold equivalent stages in a wild-type embryo, respectively) embryos of wild-type (early $n=12$, mid $n=19$, late $n=16$), *pak-1(tm403)* (early $n=16$, mid $n=21$, late $n=16$), *spc-1(RNAi)* (early $n=15$, mid $n=21$, late $n=20$), and *spc-1(RNAi) pak-1(tm403)* (early $n=12$, mid $n=17$, late $n=26$) genotypes.

(b) Actin filament orientation based on Fast Fourier Transform and binarisation. Wild type (early $n=12$, mid $n=18$, late $n=14$), *pak-1(tm403)* (early $n=16$, mid $n=20$, late $n=16$), *spc-1(RNAi)* (early $n=14$, mid $n=18$, late $n=18$), and *pak-1(tm403) + spc-1(RNAi)* (early $n=12$, mid $n=18$, late $n=21$).

Note that the characteristics of actin filaments in *spc-1(RNAi) pak-1(tm403)* embryos differ mostly at the equivalent of the two-fold stage when muscles become active. At earlier and later stages, *spc-1(RNAi)* embryos and *spc-1(RNAi) pak-1(tm403)* embryos become similar.

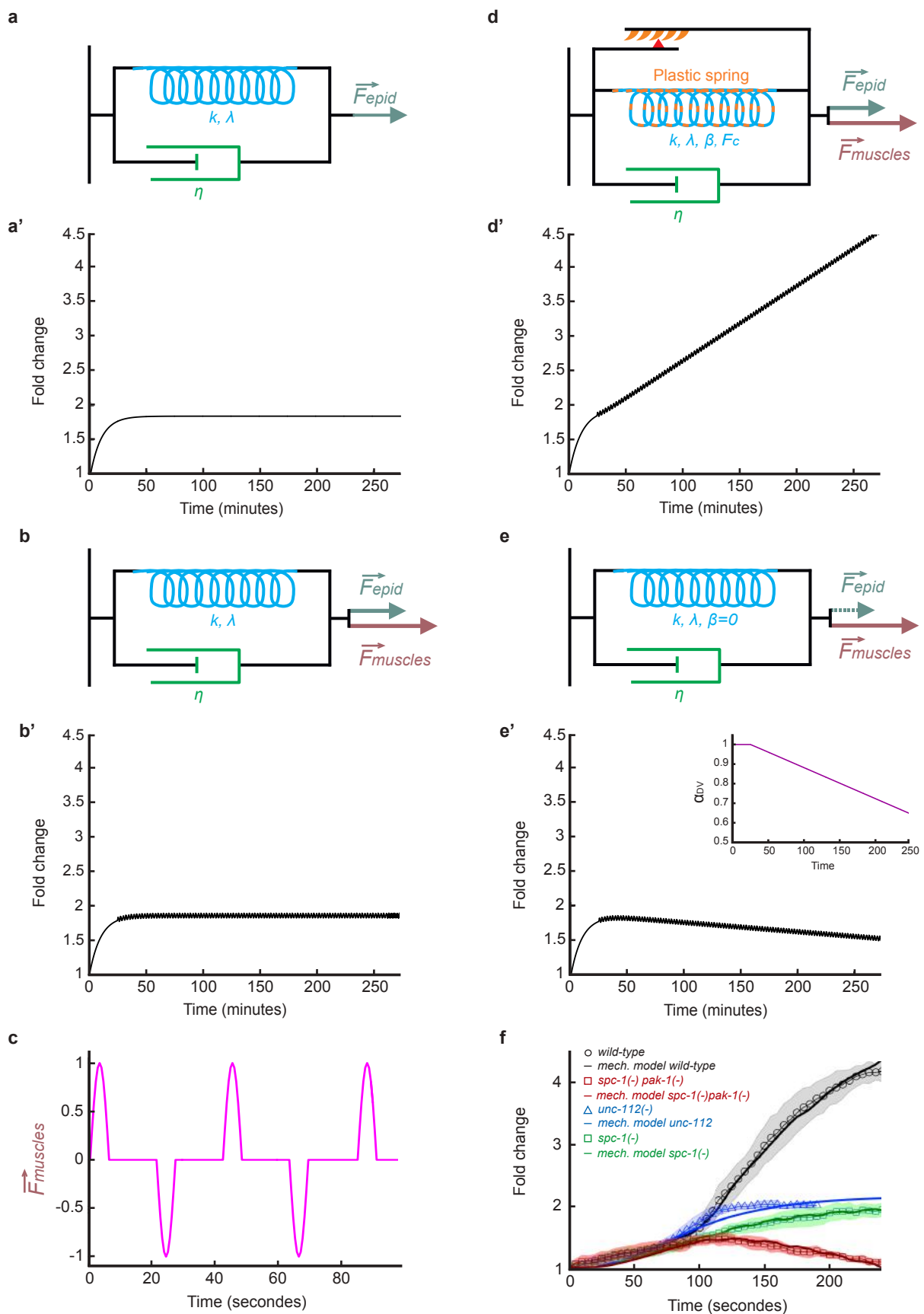
(c-f) Epidermal actin filaments visualized with the *Pepid::Lifeact::GFP* construct in wild-type (C), *spc-1(RNAi) pak-1(tm403)* (D) embryos (panels are from Fig. 3). * $<0,05$; ** $<0,001$; *** $<0,0001$; ns, not significant.



Supplementary Figure 5: Change in embryo diameter during elongation

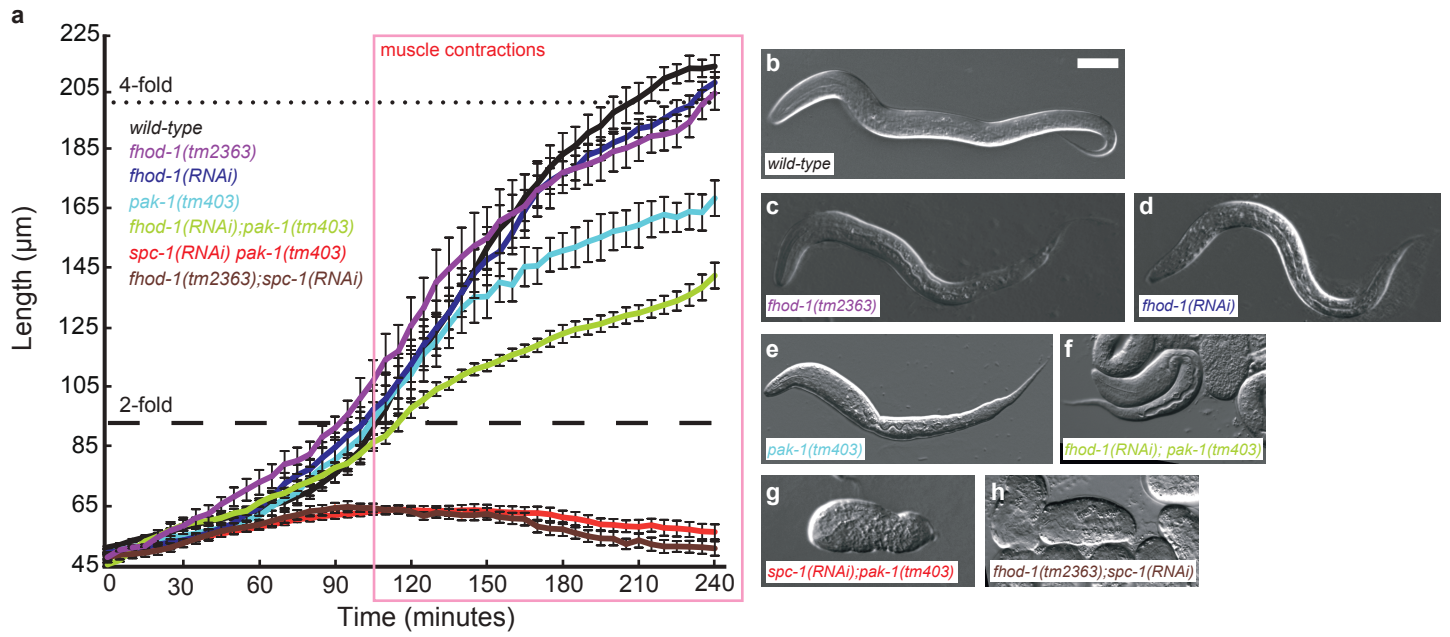
(a-b'') Fluorescence micrographs of embryos expressing the *Pepid::Lifeact::GFP* construct in the epidermis at three elongation stages early, middle and late (corresponding to 1.7-fold, 2-fold and 3-fold equivalent stages in a wild-type embryo, respectively) for wild-type (a-a'') and *spc-1(RNAi) pak-1(tm403)* embryos (b-b''); the *Pepid* promoter corresponds to *Pdpy-7*. The yellow lines correspond to the segments used to measure the dorso-ventral width of the V1 seam cell. (c) Quantification of the average V1 cell width normalized to the initial width during elongation in four genotypes. (d) Quantification of the average dorso-ventral width at the level of the V1 seam cell, which was calculated using the measured embryo length and V1 cell width, taking into consideration the conservation of the total embryo volume. Each point in panel (c) and (d) represents between 7 and 21 embryos; error bars are standard errors.

A notable feature of *spc-1(RNAi) pak-1(tm403)* embryos is that the circumferential dimension of the seam cells decreased much more than that of their DV cell, which most likely reflects the actin filament integrity defects combined with a *Fseam* force largely unchanged.



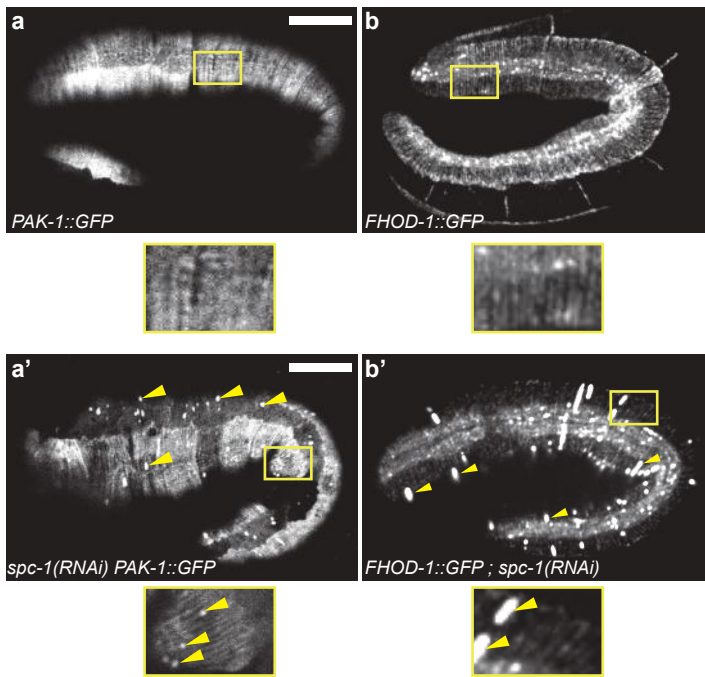
Supplementary Figure 6: Time-dependent length of a Kelvin-Voigt model in different conditions.

(a-a') A generic Kelvin-Voigt system subject to a constant force F_{epid} and its predicted elongation change using $F_{epid}=0.85$ (A').
 (b-b') A similar system subject to two forces, F_{epid} and an oscillating force $F_{muscles}$ with the properties depicted in (E), and predicted elongation change using $F_{epid}=0.85$ (B'). As the pulsatile force induces both compression and stretching (see Fig. S6), its net input on elongation is transient and the system oscillates around the maximal value reached without $F_{muscles}$.
 (d-d') A Kelvin-Voigt system with mechanical plasticity introduced according to Eqs. (1,4-7), and predicted elongation change using $F_{epid}=0.85$, $\beta=0.10$ and $F_c=0$ according to Eqs. (1,4-7). (e) A Kelvin-Voigt system in which the plasticity is defective ($\beta=0$), and there is actin tearing according to Eq. (7) inducing a progressive reduction of F_{epid} , and predicted elongation change using $F_{epid}=0.85$, with an initial value of the passive component $\alpha DV(t=0)=1$ and the tearing factor $\gamma=0.15$ (D').
 (e' Inset) Behavior of $\alpha DV(t)$.
 (c) In all conditions $F_{muscles}$ is a periodic function with positive and negative steps of duration of 6 seconds alternating with periods of null value of duration 15 seconds modulated by a cosine function. In (a-e') the elastic constant of the spring is $k=1$, the initial resting length has the value $\lambda(t=0)=1$, and the viscosity value is $\eta=10$. (f) Result of the fit for the following genotypes: wt, *spc-1 pak-1*, *unc-112* and *spc-1* according to Eqs. (1,4-7). The values of the parameters are specified in paragraphs 1.4, 1.5 and 1.6.



Supplementary Figure 7: Comparable retraction phenotypes after the combined loss of SPC-1 and PAK-1 or SPC-1 and FHOD-1

(a) Elongation curves of the single and double deficient embryos indicated on the left, (b-h) and corresponding terminal phenotypes at hatching: wild-type (n=5), *fhod-1(tm2363)* (n=6) *fhod-1(RNAi)* (n=7) *pak-1(tm403)* (n=5) *fhod-1(RNAi); pak1(tm403)* (n=7), *spc-1(RNAi) pak-1(tm403)* (n=8), *fhod-1(tm2363);spc-1(RNAi)* (n=9). Scale bar, 25 μm .

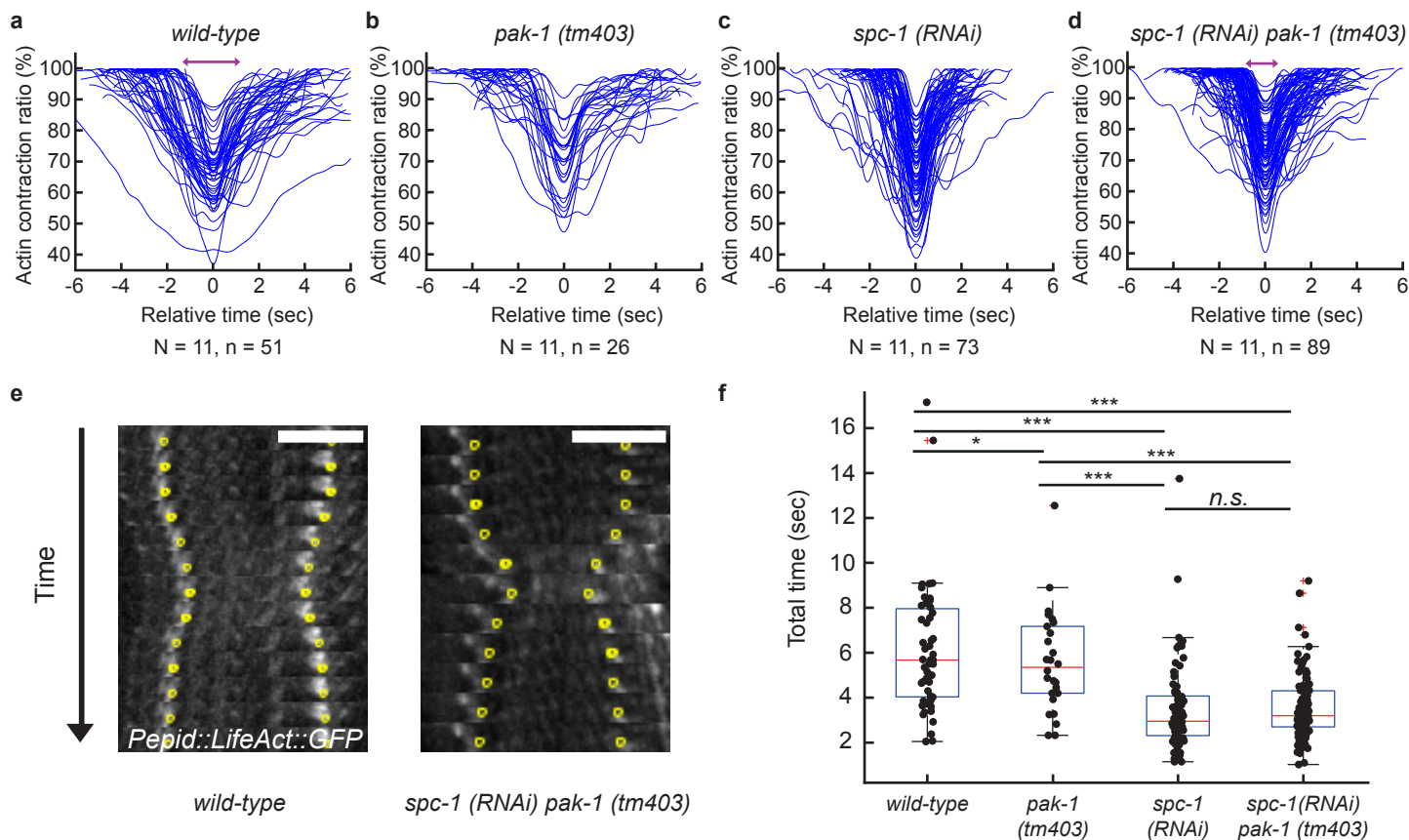


Supplementary Figure 8: PAK-1 and FHOD-1 form aggregates in *spc-1(RNAi)* loss of function.

(a-a') PAK-1::GFP localisation in wild-type and *spc-1(RNAi)* embryos. Yellow box, area enlarged below the panel. Note the punctae in SPC-1 deficient embryos.

(b-b') FHOD-1 localization in wild-type and *spc-1(RNAi)* embryos.

Note the aggregates (arrowheads). Note also that FHOD-1::GFP displayed a filamentous organization reminiscent of actin filaments. Scale bar: 10 μ m.



Supplementary Figure 9: Actin displacement ratio

(a-d) Individual displacement tracks of actin filaments visualized with a *Pdpy-7::Lifeact::GFP* marker specifically expressed in the epidermis of wild type (a), *pak-1(tm403)* (b), *spc-1(RNAi)* (c) and *spc-1(RNAi) pak-1(tm403)* (d) embryos at a stage equivalent to 2-fold in a wild-type embryo. Scale bar: 10 μ m.

(e) Typical kymographs of *Pepid::Lifeact::GFP*-labeled actin filaments in wild-type and *spc-1(RNAi) pak-1(tm403)* embryos from which the tracks in a-d were derived. Yellow dots correspond to landmarks for quantitative analysis.

(f) Quantification of the displacement duration in (N=embryo/ n=contraction): wild-type, N=11/n=51; *pak-1(tm403)*, N=11/n=26; *spc-1(RNAi)*, N=11/n=73; *spc-1(RNAi) pak-1(tm403)*, N=11/n=89. * $<0,05$; ** $<0,001$; *** $<0,0001$; ns, not significant.

A	Reproducible 50-100% enhanced defect* compared to wild-type
B	Reproducible 20-50% enhanced defect* compared to wild-type
C	Reproducible 5-20% enhanced defect* compared to wild-type

Targeted gene	Function	Strength of interaction
(III) <i>ani-1</i>	Anilin	A
(IV) <i>cap-1</i>	F-actin capping protein α subunit	A
(II) <i>cap-2</i>	β subunit of actin capping protein	A
(V) <i>cdc-25.2</i>	Putative homolog of Cdc25 phosphatase	A
(II) <i>cdc-42</i>	RHO GTPase	A
(III) <i>dlc-1</i>	Dynein light chain	A
(IV) <i>epi-1</i>	Laminin α chain	A
(I) <i>hmp-2</i>	β -catenin	A
(IV) <i>lam-1</i>	Laminin β	A
(I) <i>let-502</i>	Rho-binding Ser/Thr kinase	A
(III) <i>mhc-5</i>	Myosin II essential light chain ortholog	A
(I) <i>pfn-1</i>	Profilin	A
(V) <i>sma-1</i>	β H-spectrin	A
(X) <i>spc-1</i>	α -spectrin	A
(I) <i>sur-6</i>	Regulatory subunit of serine/threonine protein phosphatase 2A	A
(X) <i>tmi-1</i>	Troponin	A
(II) <i>dsh-2</i>	Dishevelled (Dsh) homolog	B
(I) <i>goa-1</i>	Ortholog of the heterotrimeric G protein α subunit Go	B
(I) <i>hmr-1</i>	Cadherin	B
(I) <i>kin-10</i>	Putative regulatory (β) subunit of casein kinase II	B
(V) <i>mom-2</i>	Member of the Wnt family	B
(II) <i>mpk-2</i>	Mitogen activated protein (MAP) kinase	B
(I) <i>pfd-3</i>	Putative prefoldin, orthologous to human VBP1	B
(II) <i>aakg-5</i>	AMP kinase	C
(V) <i>arx-2</i>	Subunit of the actin related protein of the conserved Arp2/3 complex	C
(III) <i>arx-3</i>	Subunit of the actin related protein of the conserved Arp2/3 complex	C
(I) <i>bub-1</i>	Serine/threonine kinase	C
(V) <i>chk-1</i>	CHK1-like serine threonin protein kinase	C
(I) <i>chp-1</i>	Protein containing two CHORD domains	C
(I) <i>csnk-1</i>	Ortholog of human CSNK1G3, CSNK1G1 and CSNK1G2	C
(II) <i>ect-2</i>	Putative RHO guanine nucleotide exchange factor (RhoGEF)	C
(I) <i>ekl-1</i>	Ortholog of members of the human TDRD	C
(IV) <i>fln-1</i>	Filamin	C
(X) <i>ifa-3</i>	Intermediate filament protein	C
(III) <i>klp-19</i>	Plus-end-directed microtubule motor protein	C
(III) <i>lit-1</i>	Serine threonine protein kinase	C
(I) <i>mei-2</i>	Novel protein containing a region similar to the p80-targeting subunit of katanin	C
(V) <i>mrck-1</i>	Serine/threonine-protein kinase	C
(III) <i>par-2</i>	Protein containing a C3HC4-type RING-finger	C
(III) <i>pfd-5</i>	Putative prefoldin 5 subunit	C
(I) <i>sys-1</i>	Novel protein that contains three divergent armadillo repeats	C
(II) <i>spv-1</i>	Ortholog of human GMIP	C
(I) <i>tbcd-1</i>	Putative β -tubulin folding cofactor D	C
(I) <i>usp-5</i>	Ortholog of human USP5 and USP13	C
(II) <i>Y19D2B.1</i>	Structural constituent of cytoskeleton	C

* Defect refers to lethality and body morphology defects

Table S1

No enhanced defect* compared to wild-type

(I) <i>afd-1</i>	(II) <i>aak-1</i>	(III) <i>abce-1</i>	(IV) <i>arp-11</i>	(V) <i>air-1</i>	(X) <i>aakb-1</i>
<i>arx-7</i>	<i>arp-1</i>	<i>abi-1</i>	<i>ced-5</i>	<i>cct-7</i>	<i>aakg-2</i>
<i>aspm-1</i>	<i>C27H5.4</i>	<i>arf-1.2</i>	<i>dli-1</i>	<i>F14H3.12</i>	<i>abl-1</i>
<i>chs-1</i>	<i>cacn-1</i>	<i>cct-5</i>	<i>dnc-1</i>	<i>gck-2</i>	<i>dyn-1</i>
<i>col-53</i>	<i>cal-2</i>	<i>cct-6</i>	<i>dyci-1</i>	<i>knl-3</i>	<i>efn-3</i>
<i>cpn-1</i>	<i>ccm-3</i>	<i>cls-1</i>	<i>eps-8</i>	<i>mig-6</i>	<i>F20B6.1</i>
<i>cutl-13</i>	<i>cct-1</i>	<i>cls-2</i>	<i>frk-1</i>	<i>noca-1</i>	<i>frm-9</i>
<i>dhc-1</i>	<i>cct-2</i>	<i>col-94</i>	<i>gex-2</i>	<i>pak-2</i>	<i>hpk-1</i>
<i>dlc-6</i>	<i>cct-4</i>	<i>col-97</i>	<i>gex-3</i>	<i>par-1</i>	<i>ifa-2</i>
<i>eak-6</i>	<i>cdc-25.4</i>	<i>cra-1</i>	<i>klp-10</i>	<i>rbx-1</i>	<i>kin-29</i>
<i>egg-5</i>	<i>cpn-2</i>	<i>cyk-4</i>	<i>klp-11</i>	<i>spas-1</i>	<i>lam-2</i>
<i>egg-6</i>	<i>dep-1</i>	<i>daf-4</i>	<i>klp-5</i>	<i>sun-1</i>	<i>lin-18</i>
<i>ekl-4</i>	<i>ebp-2</i>	<i>fem-2</i>	<i>let-60</i>	<i>syx-5</i>	<i>lpr-3</i>
<i>erm-1</i>	<i>eff-1</i>	<i>frm-2</i>	<i>let-92</i>	<i>unc-112</i>	<i>nck-1</i>
<i>fhod-1</i>	<i>egg-3</i>	<i>gei-4</i>	<i>M116.5</i>	<i>unc-70</i>	<i>pak-1</i>
<i>gei-17</i>	<i>evl-20</i>	<i>gop-3</i>	<i>nsp-1</i>		<i>pfn-2</i>
<i>gfi-2</i>	<i>F59A6.5</i>	<i>ina-1</i>	<i>par-5</i>		<i>pfn-3</i>
<i>gsa-1</i>	<i>frm-5.2</i>	<i>inf-1</i>	<i>pdf-1</i>		<i>pqn-34</i>
<i>gsk-3</i>	<i>glb-12</i>	<i>kin-18</i>	<i>pld-1</i>		<i>unc-97</i>
<i>gsp-3</i>	<i>gpb-1</i>	<i>klp-6</i>	<i>pmk-2</i>		
<i>gsp-4</i>	<i>klp-1 / unc-104</i>	<i>klp-7</i>	<i>pmk-3</i>		
<i>kca-1</i>	<i>klp-17</i>	<i>let-805</i>	<i>ptp-4</i>		
<i>lim-9</i>	<i>let-268</i>	<i>mpk-1</i>	<i>rac-1 / ced-10</i>		
<i>mel-26</i>	<i>lrr-1</i>	<i>mtm-3</i>	<i>rack-1</i>		
<i>mfap-1</i>	<i>max-2</i>	<i>nfm-1</i>	<i>unc-33</i>		
<i>mom-5</i>	<i>mel-11</i>	<i>pef-1</i>	<i>wsp-1</i>		
<i>nab-1</i>	<i>mig-5</i>	<i>plk-1</i>	<i>zen-4</i>		
<i>ned-8</i>	<i>mlt-8</i>	<i>pph-6</i>			
<i>nkb-1</i>	<i>nsy-1</i>	<i>ptp-1</i>			
<i>nmy-2</i>	<i>pdf-2</i>	<i>pxl-1</i>			
<i>npp-4</i>	<i>pink-1</i>	<i>rfl-1</i>			
<i>ocr1-1</i>	<i>pir-1</i>	<i>tbb-2</i>			
<i>pes-7</i>	<i>ptc-3</i>	<i>ten-1</i>			
<i>pdf-6</i>	<i>ptp-2</i>	<i>tlk-1</i>			
<i>ppk-1</i>	<i>ptp-3</i>	<i>trd-1</i>			
<i>rga-2</i>	<i>saps-1</i>	<i>unc-116</i>			
<i>rsa-1</i>	<i>scpl-2</i>	<i>wrm-1</i>			
<i>smgl-1</i>	<i>sds-22</i>				
<i>spd-1</i>	<i>spdl-1</i>				
<i>tba-2</i>	<i>tac-1</i>				
<i>ttx-7</i>	<i>tba-4</i>				
<i>unc-35</i>	<i>unc-52</i>				
<i>unc-59</i>	<i>vab-19</i>				
<i>unc-73</i>	<i>vab-9</i>				
<i>unc-73</i>	<i>vhp-1</i>				
<i>unc-94</i>	<i>vps-11</i>				
<i>vab-10</i>	<i>vps-32</i>				
<i>viln-1</i>	<i>W0761.1</i>				
<i>vps-20</i>	<i>zyg-9</i>				
<i>wve-1</i>					

Global PBS /Predicted Biological Score/ <i>categories computed and established by Hybrigenics, to assess the interaction reliability</i>	
A	Very high confidence in the interaction
B	High confidence in the interaction
C	Good confidence in the interaction
D	Moderate confidence in the interaction <i>/either due to false-positive interactions or due to interactions that are hardly detectable by the Y2H technique/</i>

Strongest candidates
/ Prey library: C. elegans embryo /

Primary Yeast Two Hybrid Screen		
<i>/ Bait: CePAK-1 N-terminal amino acids: 1-294 /</i>		
Interactor candidate	Protein function	Global PBS
SPC-1	α -spectrin	A
F47B10.1	β -chain succinyl-co-A ligase	A
CHW-1	RhoU homolog	A
GCK-1	STE20-family kinase	A
NCK-1	NCK adaptor	A
PIX-1 *	ARHGEF7 homolog β -Pix	A
CDC-42 *	Small GTPase	A
PAK-1 *	P21-activated Ser/Tre kinase (multiple hits through kinase domain)	A
CED-10 *	Rac-1 / Small GTPase	B
POD-2	Predicted acetyl-coA carboxylase	B
Y39E4A.3	Transketolase	B
EEL-1	HECT-ubiquitin ligase	C
NPP-21	Nuclear pore protein	C
TAG-143	Transcription factor	C
UNC-44	Ankyrin	D
HIPR-1	SLA2 and Hip related	D
T05C1.4	Conserved calmodulin-binding TFs	D
Y53F4B.13	RNA methyltransferase	D
PTP-3	LAR-like receptor tyr-protein phosphatase	D
COGC-6	Conserved Oligomeric Golgi (COG) Component	D
DAF-21	Hsp90 molecular chaperone family member	D
GCK-1 variant	STE20-family kinase	D

Secondary Yeast Two Hybrid Screen		
<i>/ Bait I: CeSPC-1 SR8-10(aa:796-1243) /</i>		
<i>/ Bait II: CeSPC-1 SH3 (aa:986-1041) /</i>		
<i>/ common hits using Bait I and Bait II /</i>		
Interactor candidate	Protein function	Global PBS
PAK-1	p21-activated Kinase	A
LIM-8	LIM domain muscle component	A
F44.E2.3	ARGLU1 ortholog	A
CSN-5	COP9-subunit ortholog, E3 ubiquitin ligase interactor	A
DEB-1	Vinculin	A
DnaJ	DNAJ/ZRF1/MPP11 ortholog; ribosome-associated chaperone	B
CYLD-1	Human CYLD1 ortholog, NF- κ B signalling interactor	B
VAB-3 / VAR-1	Homeodomain protein PAX6 ortholog	B
GRL-4	Hedgehog-like protein	C
UNC-34	Enabled/VASP homolog	C
T04F8.6	Human ninein and ninein-like (GSK3B interactor) ortholog	D
ALR-1	Human ARX(aristaless) ortholog homeodomain transcription factor	D
ATN-1	α -actinin homolog	D
MMCM-1	Methylmalonyl-CoA mutase	D
UNC-70	β_6 -spectrin	D
VAB-10	Spectraplaklin	D
F26A10.2	Zinc-finger containing protein (required for germ line maintenance and locomotion)	D
F43C1.1	Human PHLPP1&PHLPP2 (PH domain&leucine- rich repeat Protein Phosphatase) ortholog	D
ALP-1	Enigma family member ALP (α -actinin associated LIM Protein) ortholog; required for actin anchorage	D
CIT-1.2	Cyclin T ortholog, embryonic transcriptional regulator	D
FLH-1	FLYWCH zinc finger transcription factor homolog	D
SHW-1	Human KCNC3 voltage-gated SHaW family potassium channel ortholog	D

* Positive controls (PAK-1 itself + its known interactors)

A	Shorter compared to <i>spc-1(-)</i>
B	Not shorter than <i>spc-1(-)</i>

Targeted gene	Function	Strength of interaction
(I) <i>fhod-1</i>	Formin	A
(I) <i>hmr-1</i>	Cadherin	A
(I) <i>hmp-2</i>	β -catenin	A
(II) <i>cdc-42</i>	RHO GTPase	A
(II) <i>spdl-1</i>	Coiled-coil protein	A
(II) <i>vps-11</i>	Ortholog of human VPS11	A
(III) <i>mtm-3</i>	Myotubularin lipid phosphatase	A
(III) <i>mhc-5</i>	Myosin II essential light chain ortholog	A
(IV) <i>dnc-1</i>	Ortholog of the dynactin complex subunit p150/GLUED/DCTN1	A
(IV) <i>epi-1</i>	Laminin α chain	A
(I) <i>goa-1</i>	Ortholog of the heterotrimeric G protein α subunit Go	B
(I) <i>kin-10</i>	Putative regulatory (β) subunit of casein kinase II	B
(I) <i>mec-8</i>	mRNA processing factor	B
(I) <i>nmy-2</i>	Non-muscle myosin II	B
(I) <i>unc-94</i>	Tropomodulin	B
(II) <i>cap-2</i>	β subunit of actin capping protein	B
(II) <i>evl-20</i>	Ortholog of human ADP-ribosylation factor-like protein 2	B
(II) <i>spv-1</i>	Ortholog of human GMIP	B
(II) <i>unc-52</i>	Perlecan	B
(III) <i>kfp-7</i>	Ortholog of human KIF2A, 2B and 2C	B
(III) <i>mup-4</i>	Transmembrane protein	B
(IV) <i>cap-1</i>	F-actin capping protein α subunit	B
(IV) <i>eps-8</i>	Cell signaling adaptor protein	B
(IV) <i>frk-1</i>	Non-receptor tyrosine kinase	B
(IV) <i>unc-33</i>	Conserved member of the CRMP/TOAD/Ulip/DRP family	B

Table S3

Name	Genotype
DM3409	<i>mnDp33 (X;IV)/+ IV.; spc-1(ra409) X.</i>
DWP10	<i>fhod-1(tm2363) I.; qals8001 [unc-119(+) fhod-1::gfp]</i>
ML1694	<i>pix-1(gk416)X.</i>
ML1725	<i>mcEX567 [spc-1::GFP, myo-2p::mcherry]</i>
ML1822	<i>sma-1(ru18)V.; pak-1 (tm403)X.</i>
ML1943	<i>mcls55[pak-1::GFP;pRF4]</i>
ML1911	<i>git-1(tm1962)X.</i>
ML2113	<i>mcls67 [dpy7p::LifeAct::GFP; unc-119(+)] V.; stls10088[hlh-1::his-24::mCherry, unc-119(+)]</i>
ML2129	<i>pak-1(tm403) X.</i>
ML2200	<i>pak-1(tm403) X.; mcls67 [dpy7p::LifeAct::GFP; unc-119(+)] V; stls10088[hlh-1::his-24::mCherry, unc-119(+)]</i>
ML2419	<i>mcEx915[ppak-1::pak-1::mkate;pR4(rol);pBSK]</i>
ML2428	<i>sma-1(ru18)V.</i>
ML2436	<i>spc-1(ra409) X.; mcEx636 [spc-1p::spc-1::GFP]</i>
ML2446	<i>pak-1(tm403) X.; spc-1(ra409) X.; mcEx636 [spc-1p::spc-1::GFP]</i>
ML2465	<i>mcls91[linc26p::ABD::mkate; myo-2p::mcherry]</i>
ML2684	<i>mcEx1008 [fhod-1 ΔFH2/DAD]</i>
ML2688	<i>pak-1(tm403) X.; mcEx1009 [fhod-1 ΔFH2/DAD]</i>
ML2853	<i>pak-1(tm403) X.; mcEx1002 [fhod-1 ΔFH1/FH2/DAD]</i>
ML2854	<i>pak-1(tm403) X.; mcEx1003 [fhod-1 ΔDAD]</i>
ML2855	<i>pak-1(tm403) X.; mcEx1004 [fhod-1 full length]</i>
ML2856	<i>mcEx1005 [fhod-1 ΔFH1/FH2/DAD]</i>
ML2857	<i>mcEx1006 [fhod-1 ΔDAD]</i>
ML2858	<i>mcEx1007 [fhod-1 full length]</i>
N2	<i>Bristol</i>
XA8001	<i>fhod-1(tm2363) I.</i>

Table S4